

Cell-based modeling of tissue-level responses to mechanical strain

Roeland Merks

Lisanne Rens, René van Oers, Hannan Tahir,
Theo Smit (AMC) and lab members

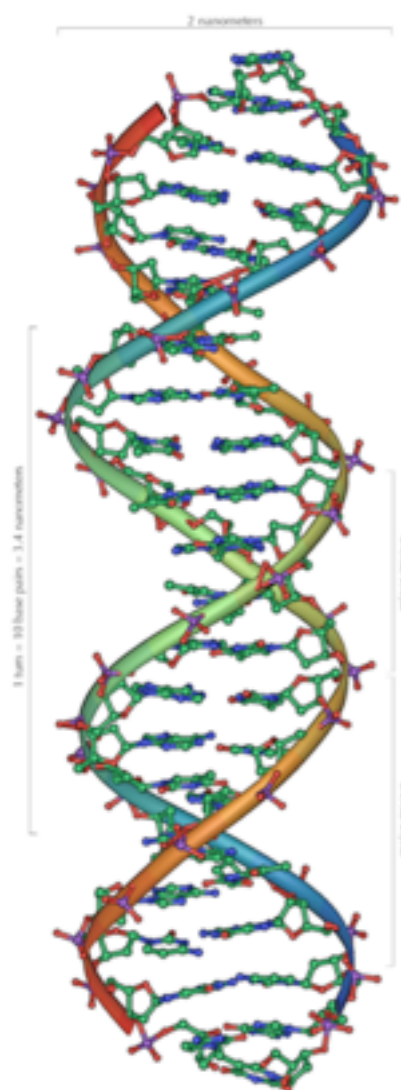
Centrum Wiskunde & Informatica (CWI), Amsterdam
Mathematical Institute, Leiden University

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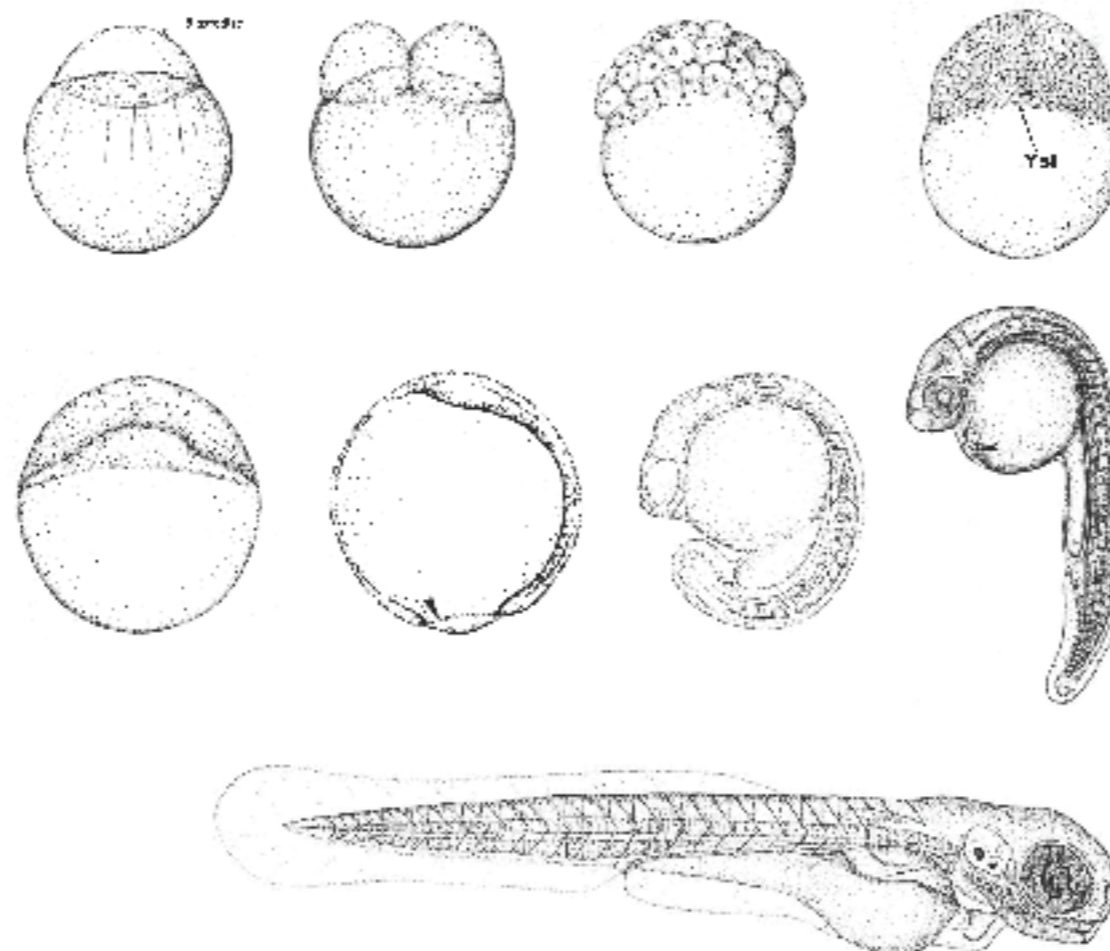
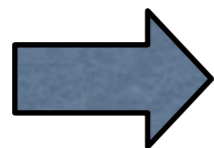
Eighth Workshop
Dynamical Systems Applied to Biology and Natural Sciences
Évora, Portugal, February 1st 2017

Morphogenesis

How is the **linear** information in the DNA translated into the **three-dimensional** shape of organisms?



DNA



Morphogenesis



Adult

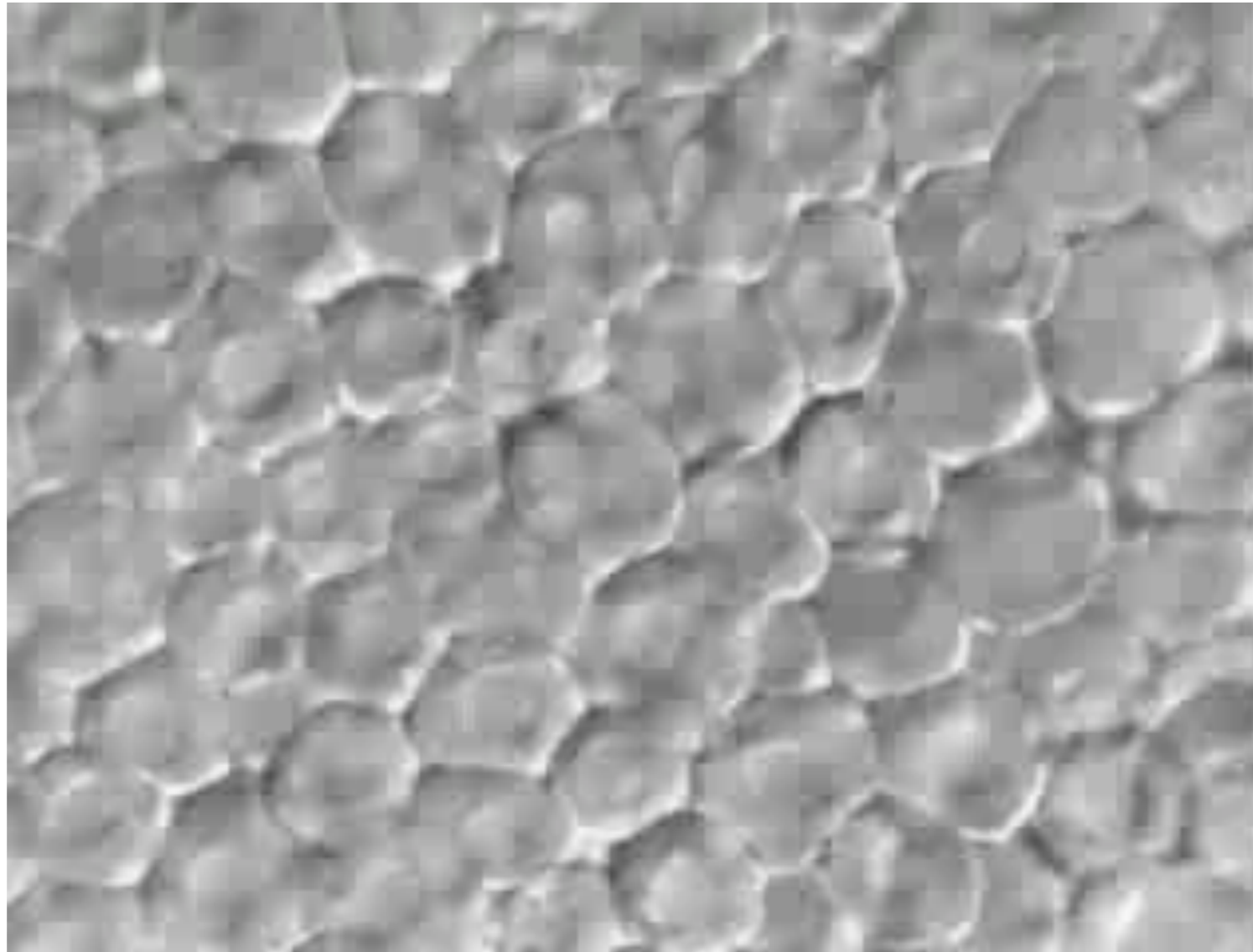
Animals are 'swarms' of cells



- Predict 'swarm' properties from cell-cell interactions
- Observe local cell-cell interactions responsible for it

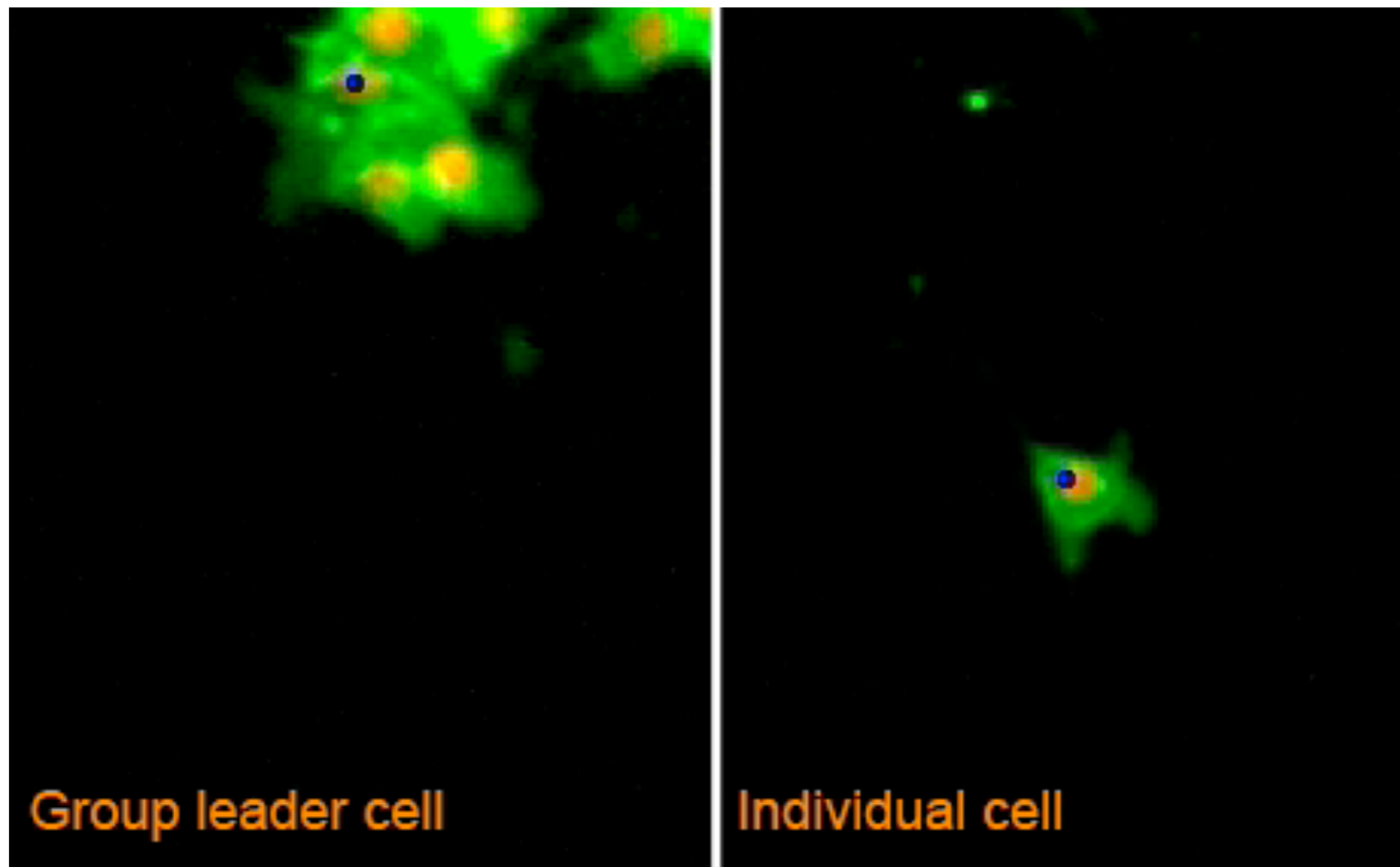
Cells look as if they act independently...

P.Z. Myers, University of Minnesota
(source: YouTube)



Zebrafish blastoderm (embryonic tissue)

... but of course cells respond to one another



Contact-inhibition of frog neural crest cells
Carmona-Fontaine *et al.* Nature 2008 (Mayer/Stern group)

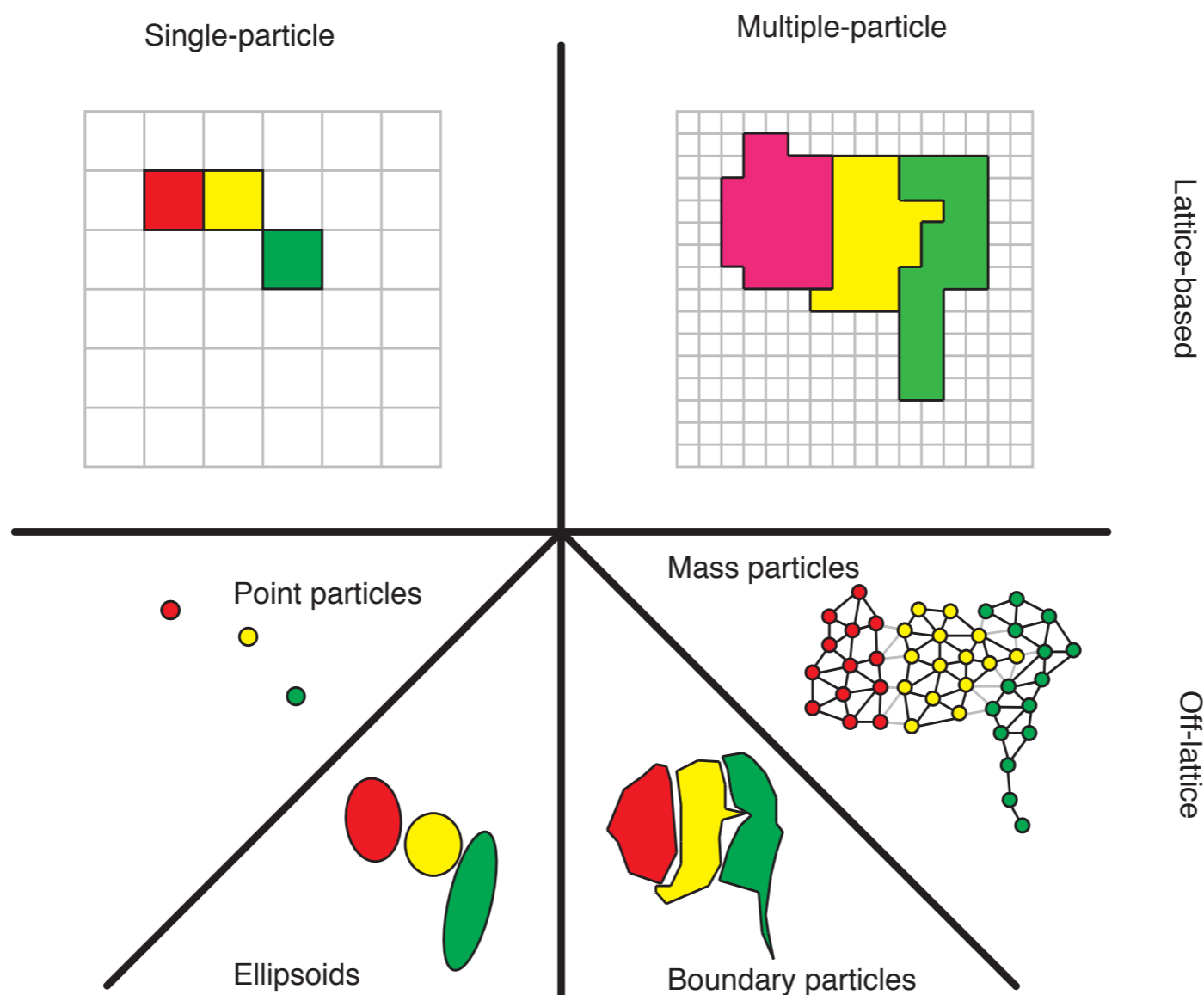
Cell based models

(Merks and Glazier, *Phys. A* 2005)

- **Input:** cell behavior
- **Output:** development of multicellular structure
 - Growth and form of tissues and organs

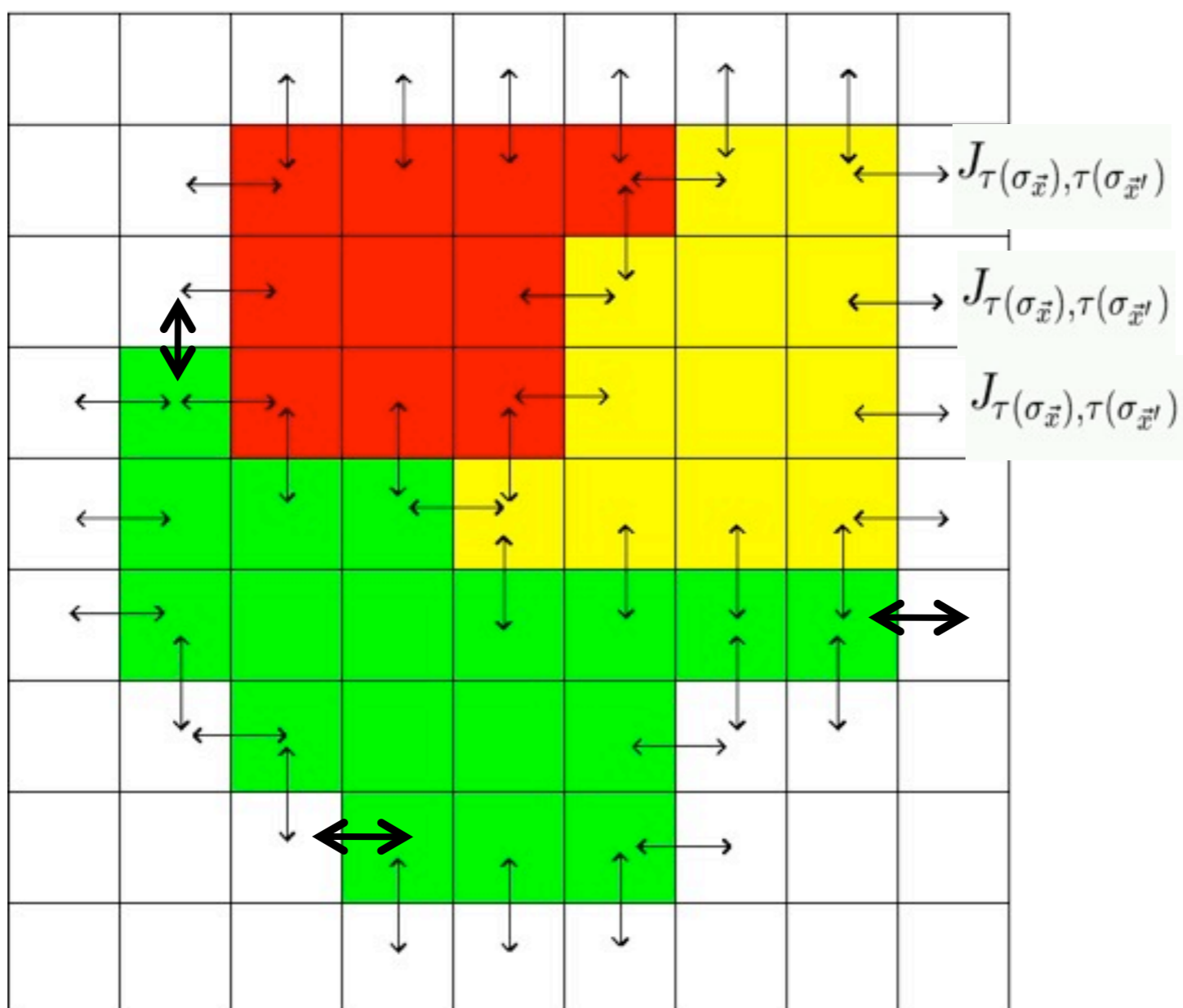
- **Aim** of cell-based modeling is to understand:
 - How cells ‘build’ animals
 - How tissue structure feeds back onto cell behavior
 - How a genetic mutation can lead to phenotypic changes

Simulation methods



- Membrane movement and cell shape are often key
 - So: **multiparticle methods**

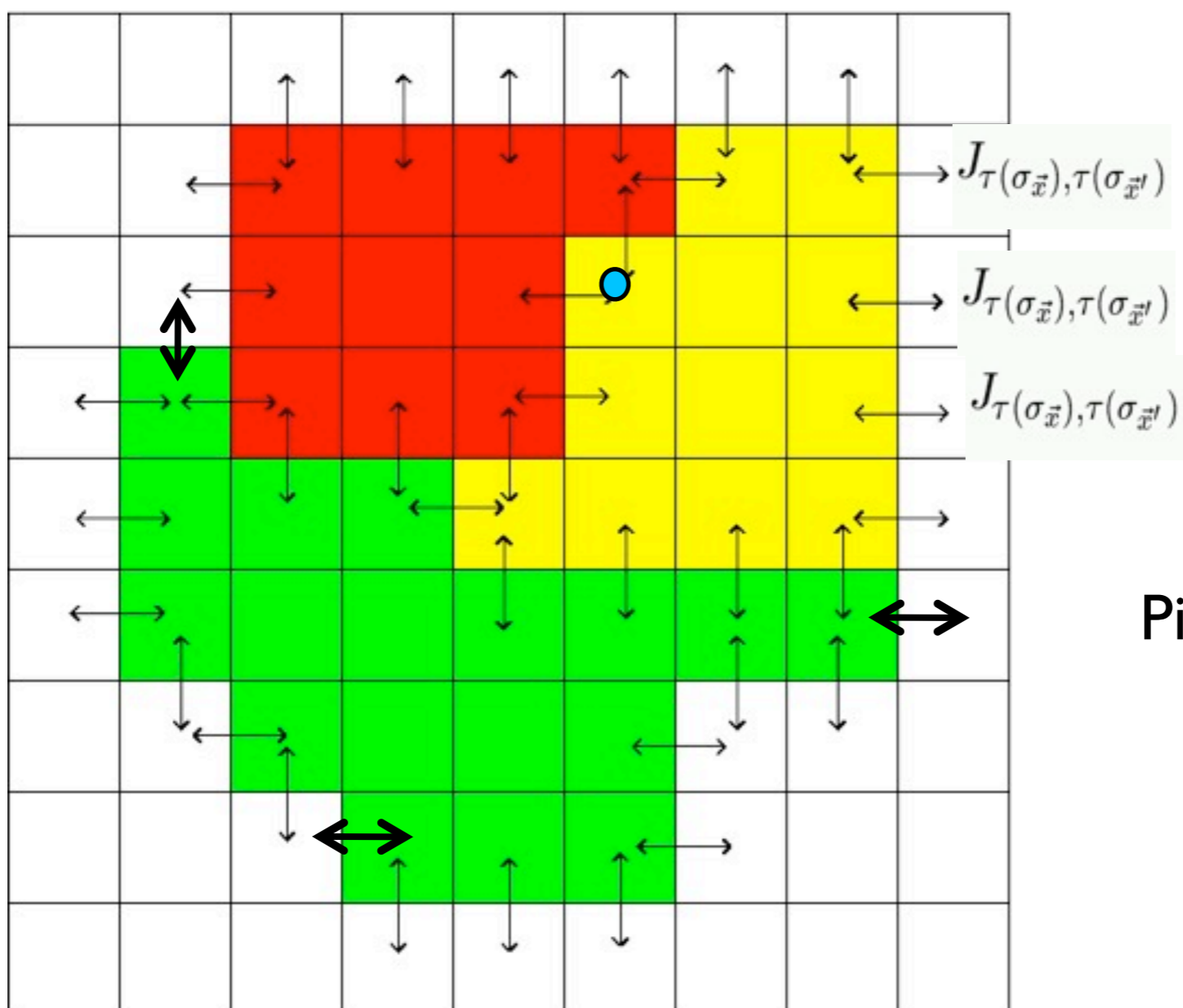
Cellular Potts Model (CPM)



$$H = \sum_{\vec{x}, \vec{x}'} J_{\tau(\sigma_{\vec{x}}), \tau(\sigma_{\vec{x}'})} (1 - \delta_{\sigma_{\vec{x}}, \sigma_{\vec{x}'}}) + \lambda \sum_{\sigma} (a_{\sigma} - A_{\sigma})^2$$

Cell adhesion
Volume conservation

Cellular Potts Model (CPM)

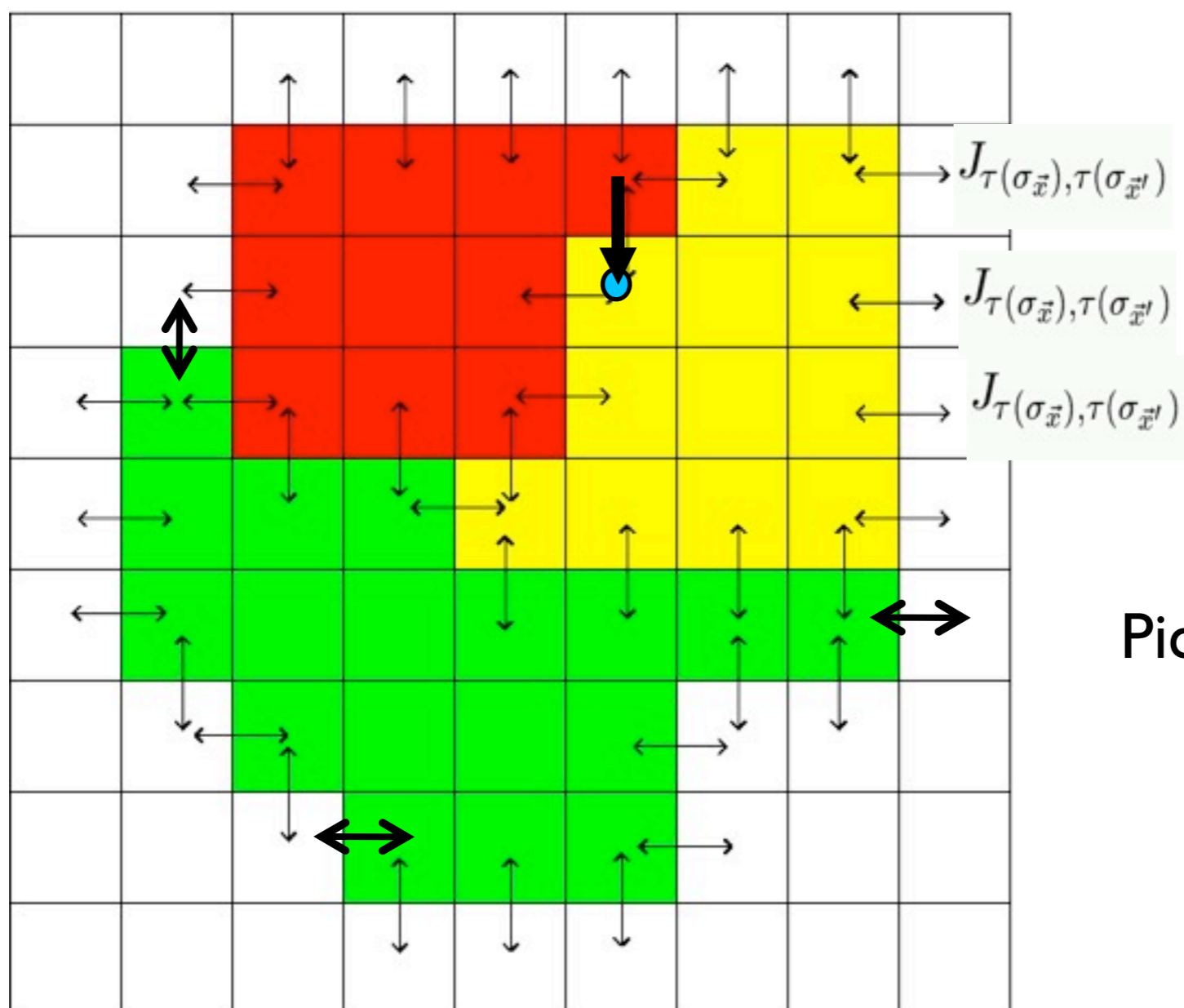


Pick random site

$$H = \sum_{\vec{x}, \vec{x}'} J_{\tau(\sigma_{\vec{x}}), \tau(\sigma_{\vec{x}'})} (1 - \delta_{\sigma_{\vec{x}}, \sigma_{\vec{x}'}}) + \lambda \sum_{\sigma} (a_{\sigma} - A_{\sigma})^2$$

Cell adhesion
Volume conservation

Cellular Potts Model (CPM)

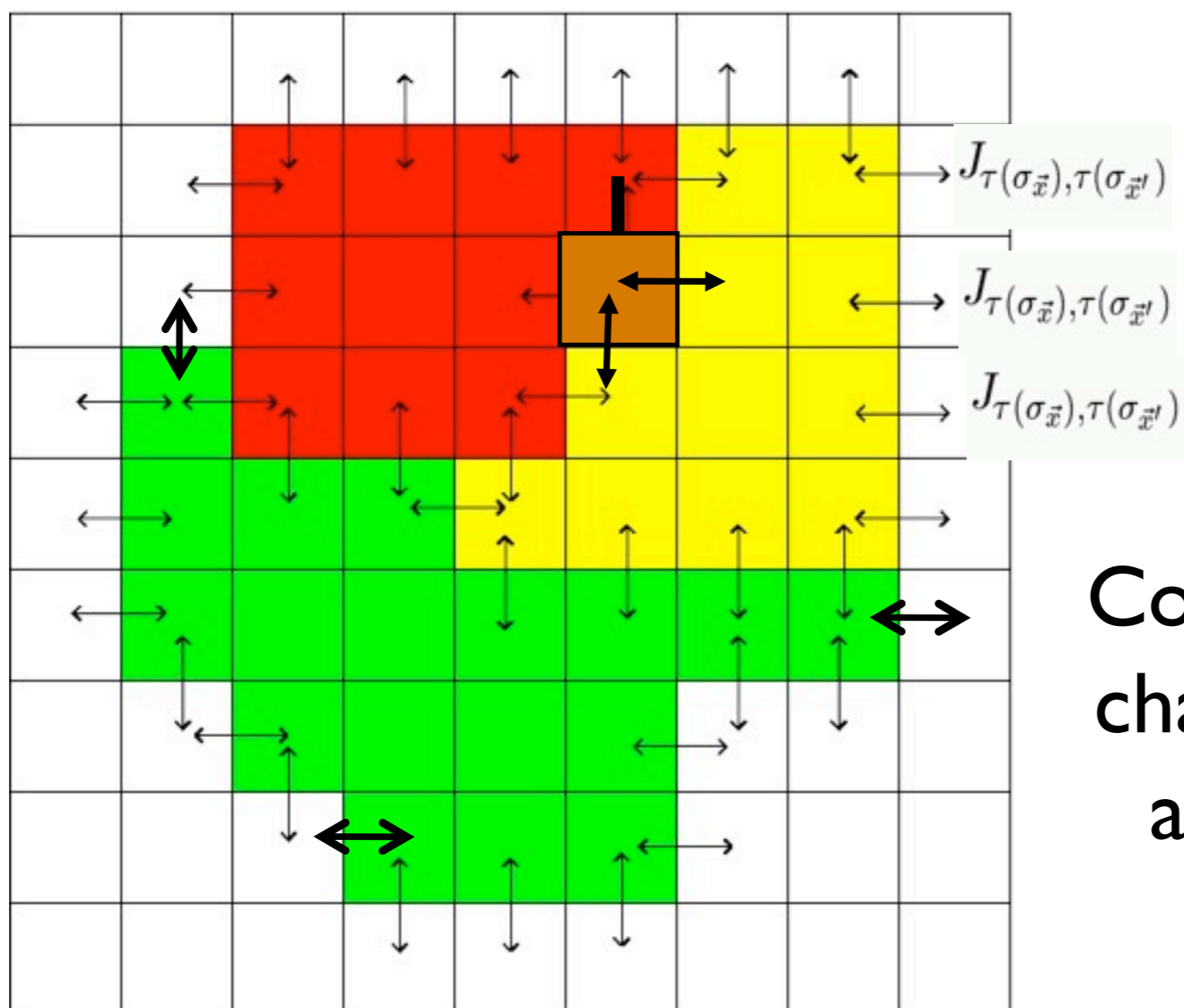


Pick random neighbor

$$H = \sum_{\vec{x}, \vec{x}'} J_{\tau(\sigma_{\vec{x}}), \tau(\sigma_{\vec{x}'})} (1 - \delta_{\sigma_{\vec{x}}, \sigma_{\vec{x}'}}) + \lambda \sum_{\sigma} (a_{\sigma} - A_{\sigma})^2$$

Cell adhesion
Volume conservation

Cellular Potts Model (CPM)

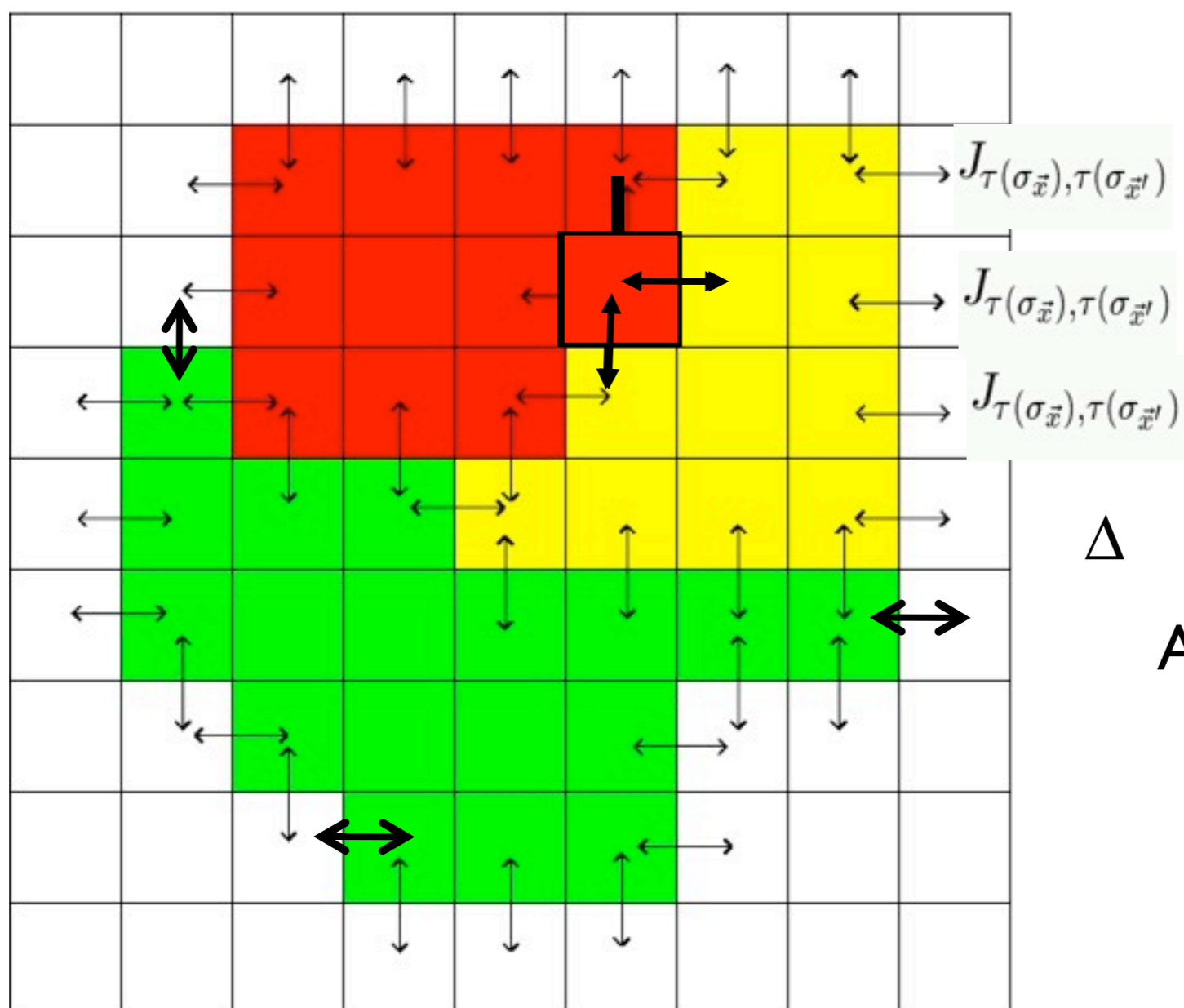


Consider energy change ΔH if we accepted this copying

$$H = \sum_{\vec{x}, \vec{x}'} J_{\tau(\sigma_{\vec{x}}), \tau(\sigma_{\vec{x}'})} (1 - \delta_{\sigma_{\vec{x}}, \sigma_{\vec{x}'}}) + \lambda \sum_{\sigma} (a_{\sigma} - A_{\sigma})^2$$

Cell adhesion Volume conservation

Cellular Potts Model (CPM)



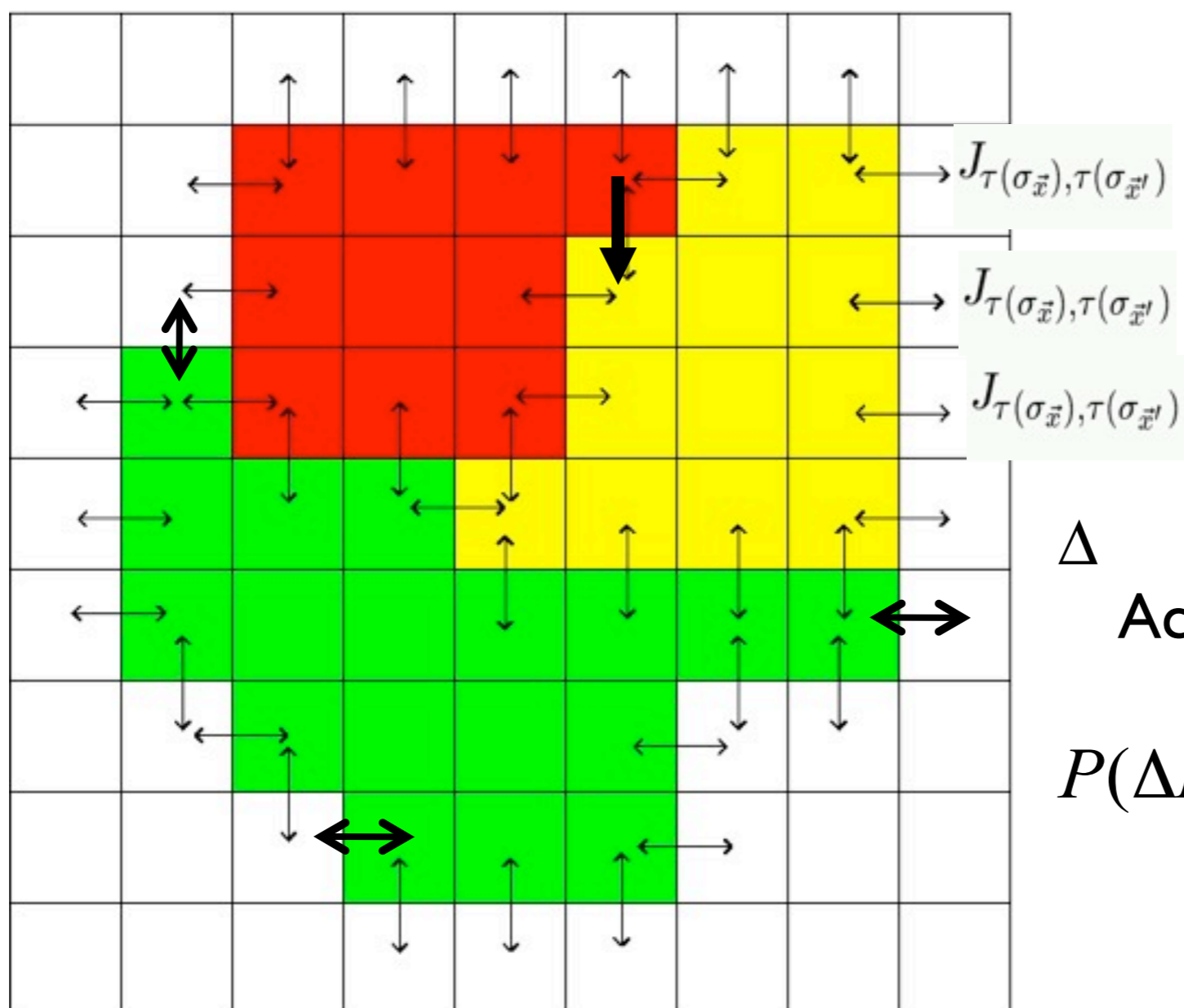
$\Delta H < 0 ?$

Accept always

$$H = \sum_{\vec{x}, \vec{x}'} J_{\tau(\sigma_{\vec{x}}), \tau(\sigma_{\vec{x}'})} (1 - \delta_{\sigma_{\vec{x}}, \sigma_{\vec{x}'}}) + \lambda \sum_{\sigma} (a_{\sigma} - A_{\sigma})^2$$

Cell adhesion
Volume conservation

Cellular Potts Model (CPM)



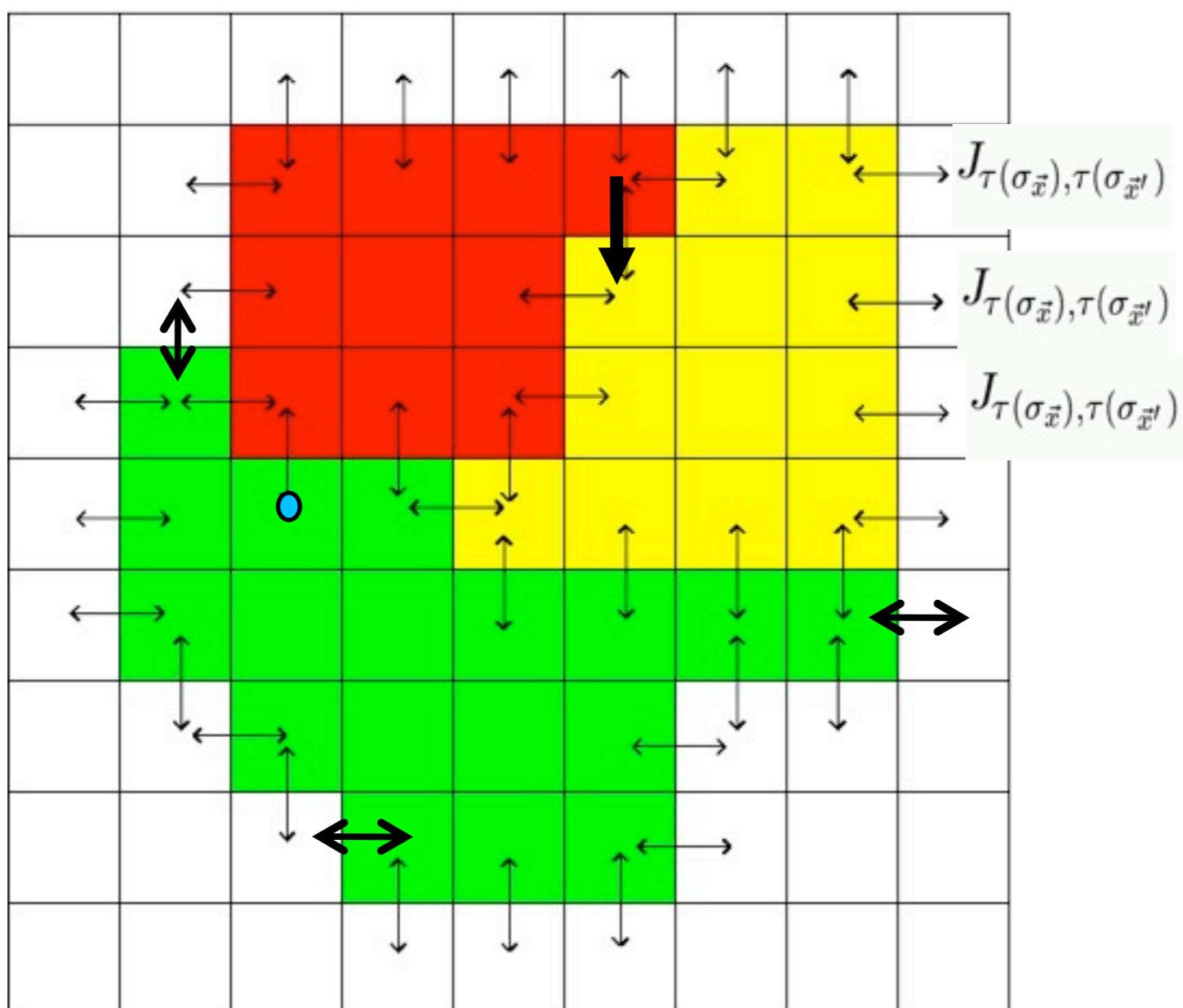
$\Delta H > 0 ?$
Accept with

$$P(\Delta H) = e^{\frac{-\Delta H + H_0}{T}}$$

$$H = \sum_{\vec{x}, \vec{x}'} J_{\tau(\sigma_{\vec{x}}), \tau(\sigma_{\vec{x}'})} (1 - \delta_{\sigma_{\vec{x}}, \sigma_{\vec{x}'}}) + \lambda \sum_{\sigma} (a_{\sigma} - A_{\sigma})^2$$

Cell adhesion Volume conservation

Cellular Potts Model (CPM)

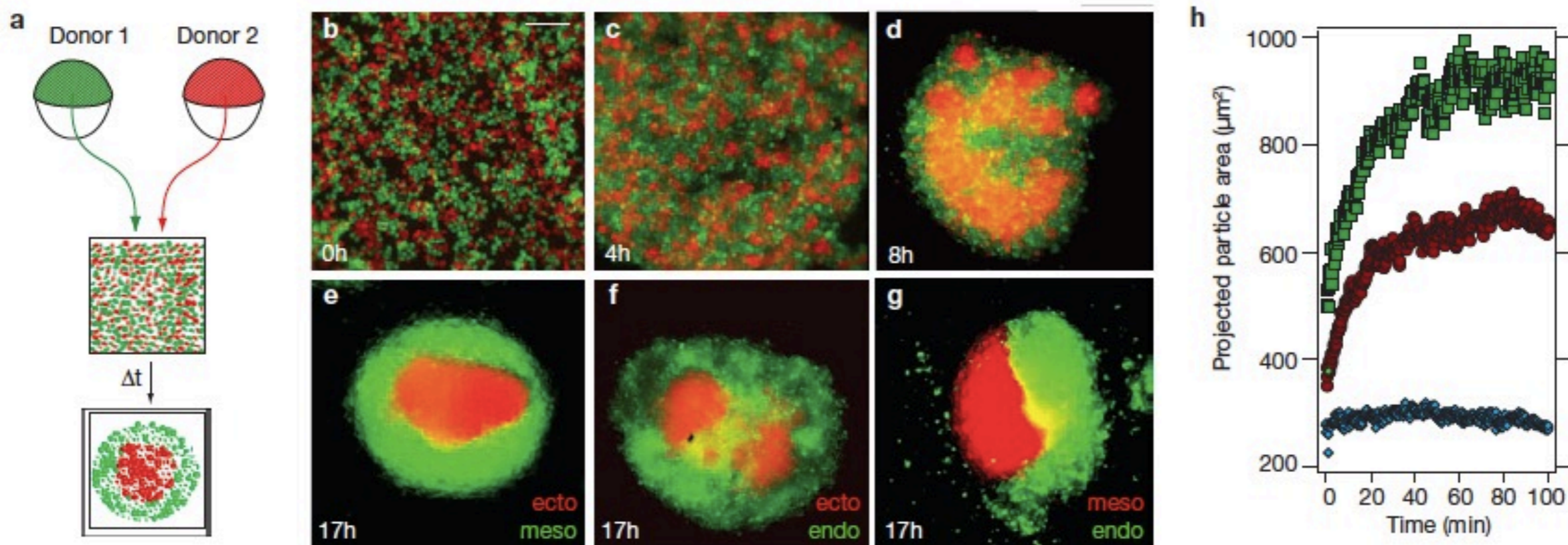


Repeat

$$H = \sum_{\vec{x}, \vec{x}'} J_{\tau(\sigma_{\vec{x}}), \tau(\sigma_{\vec{x}'})} (1 - \delta_{\sigma_{\vec{x}}, \sigma_{\vec{x}'}}) + \lambda \sum_{\sigma} (a_{\sigma} - A_{\sigma})^2$$

Cell adhesion
Volume conservation

Typical problem: Differential-adhesion-driven cell sorting



Data: Krieg et al. Nature Cell Biology, 2008

Simulation result

$$J_{\text{green,green}} < J_{\text{red,red}}$$

$$J_{\text{red,green}} = J_{\text{red,red}}$$

Modelsimulatie

10x versneld

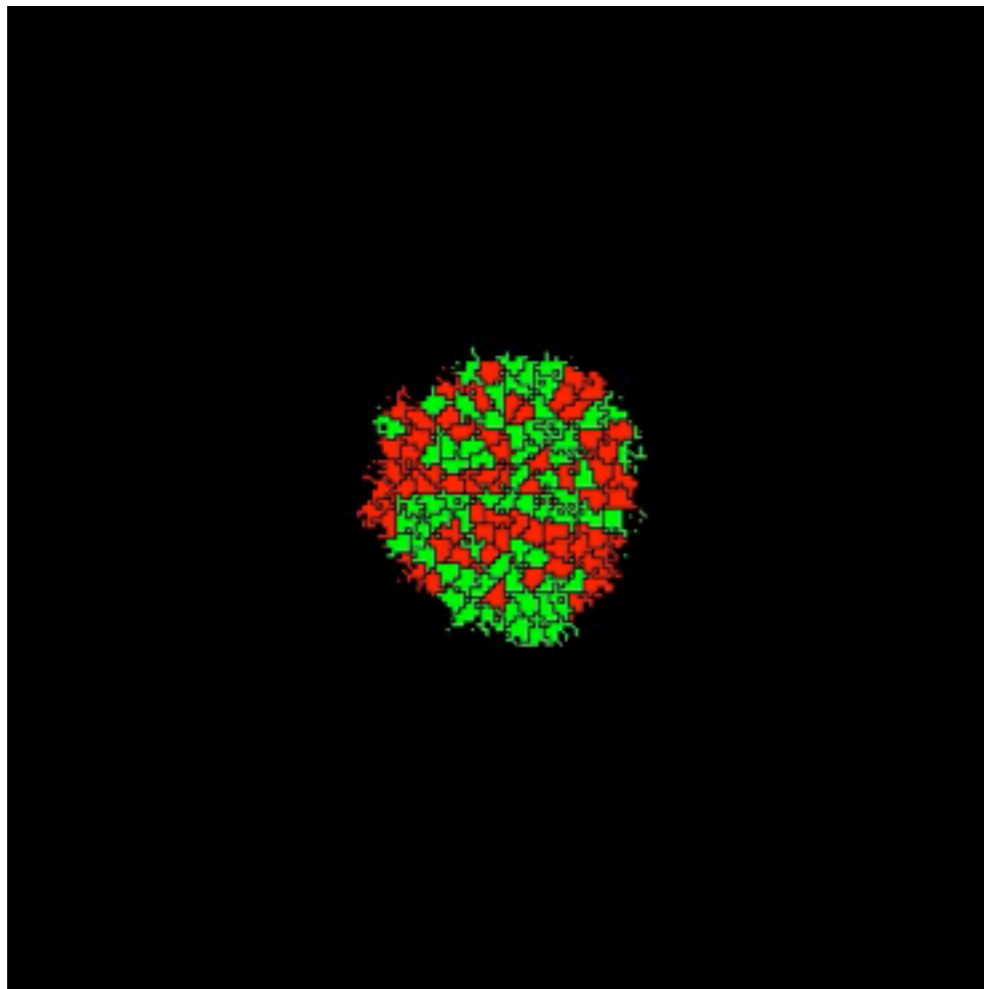
ecto
meso

Simulation result

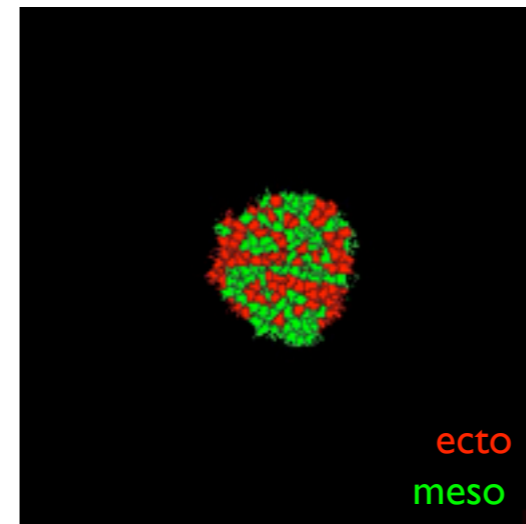
$$J_{\text{green,green}} < J_{\text{red,red}}$$

$$J_{\text{red,green}} = J_{\text{red,red}}$$

Modelsimulatie



10x versneld

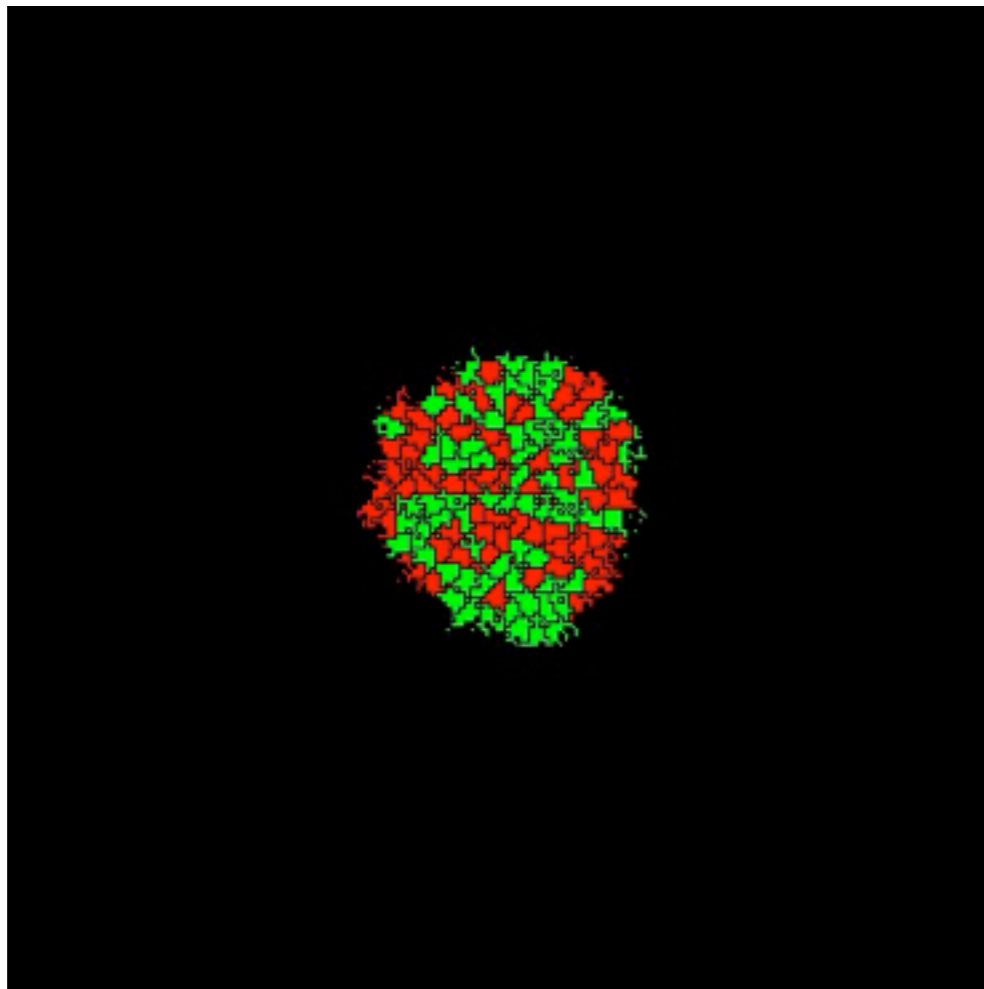


Simulation result

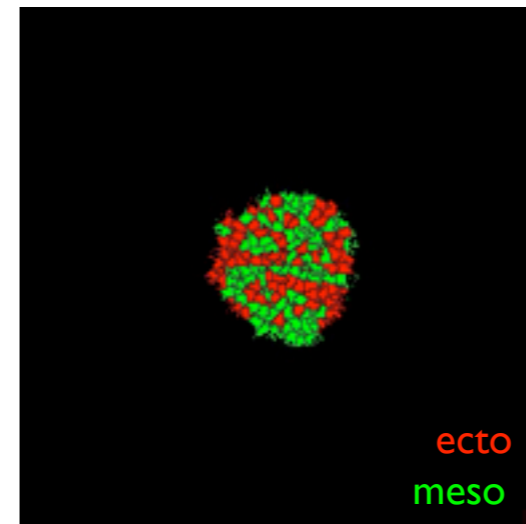
$$J_{\text{green,green}} < J_{\text{red,red}}$$

$$J_{\text{red,green}} = J_{\text{red,red}}$$

Modelsimulatie



10x versneld



Alternative representation: Boundary-element model



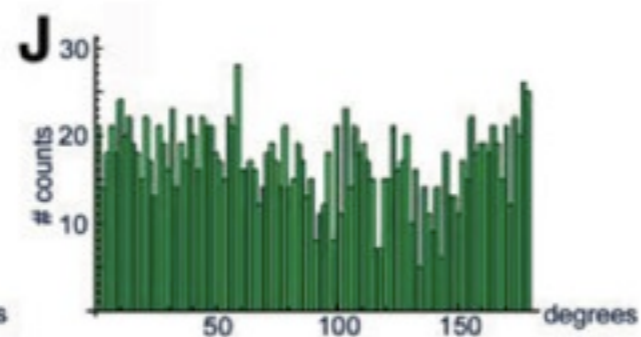
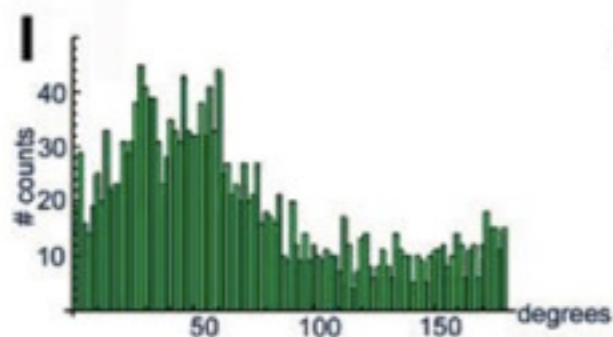
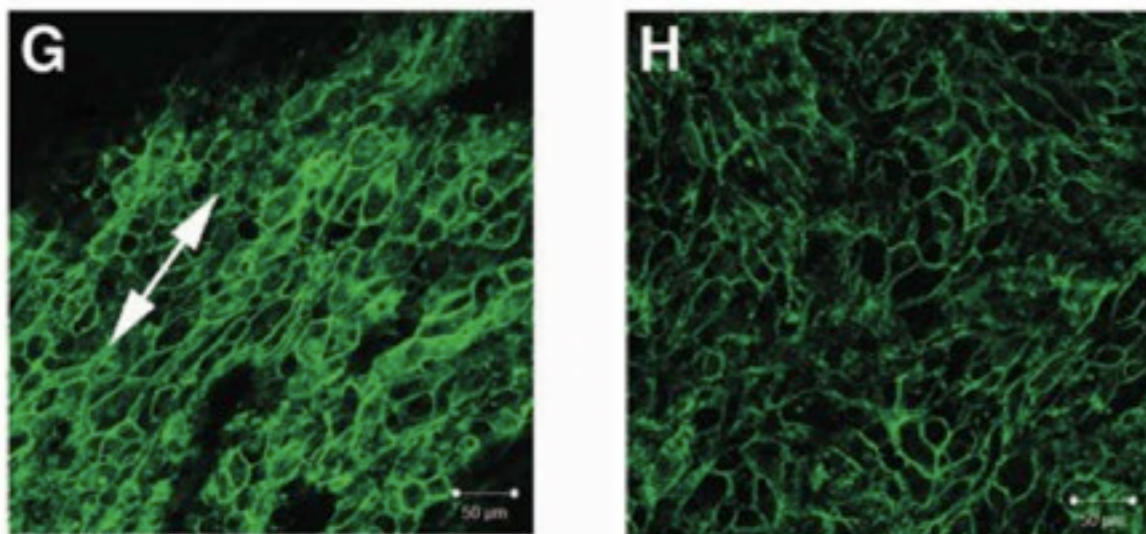
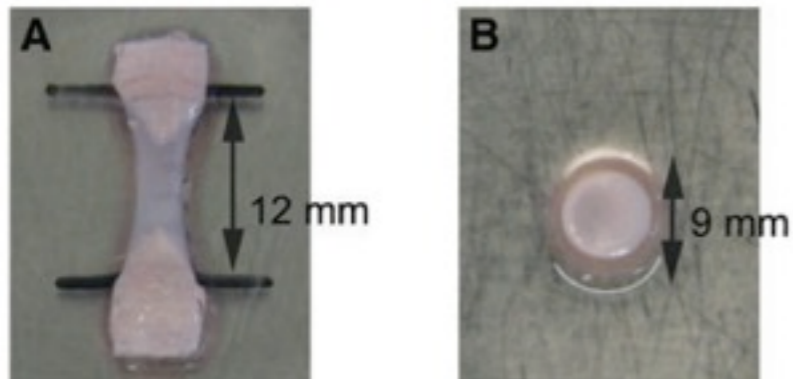
Harold Wolff

Coordination of tissue growth during morphogenesis

- If one tissue grows in the embryo:
 - adjacent tissues need to follow
 - internal structure of tissues needs to adapt to strain
- Examples:
 - Relative growth of bones and muscles
 - Muscle fibers must be oriented parallel or at a specific angle to the long axis of the muscle
 - Connective tissues and skin
 - Orientation of segments along the extending body axis

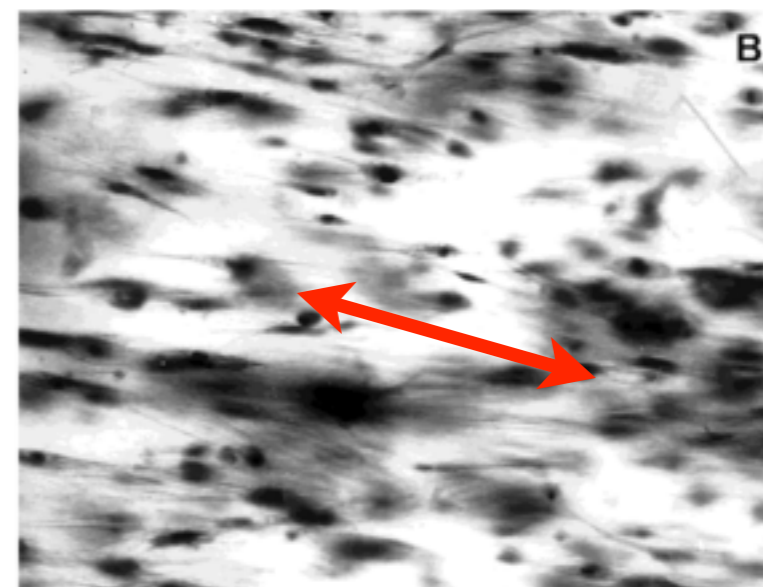
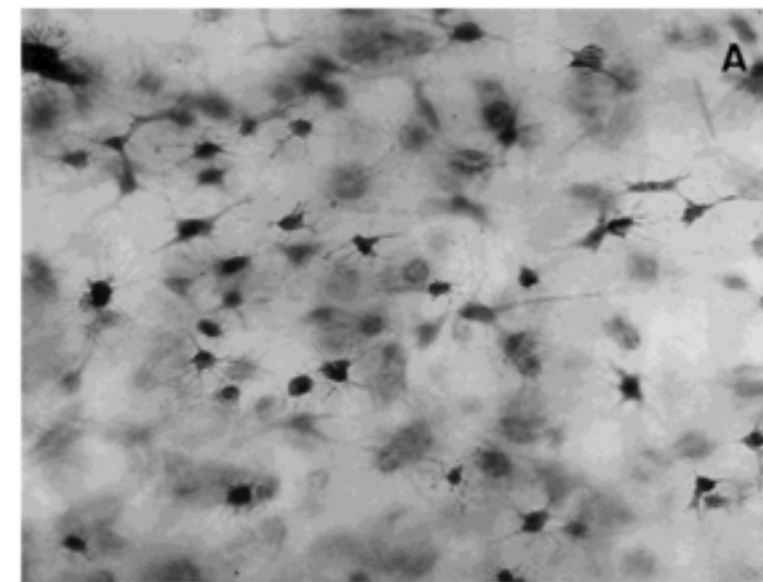
In vitro model: Cells align to static strain in fibrous substrates

Endothelial cells on collagen (BAEC)



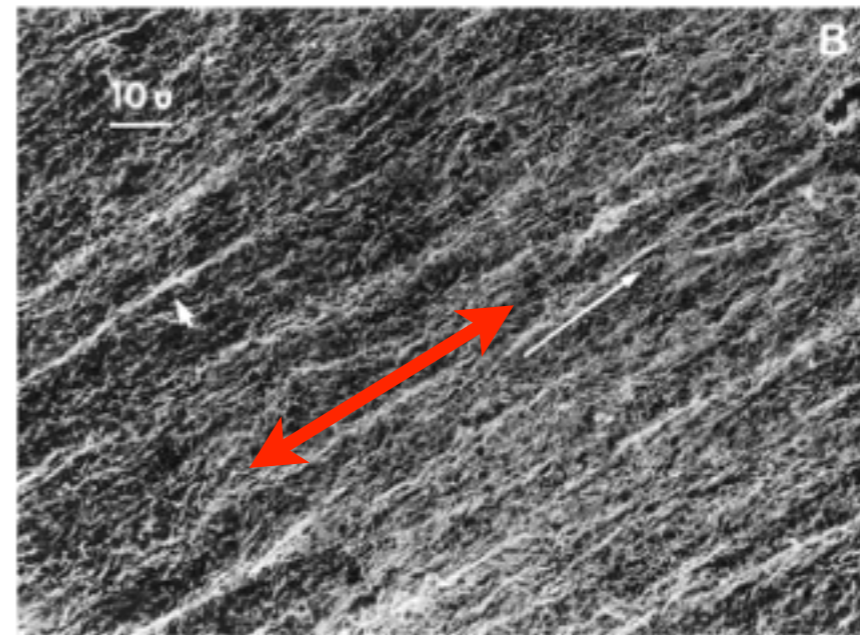
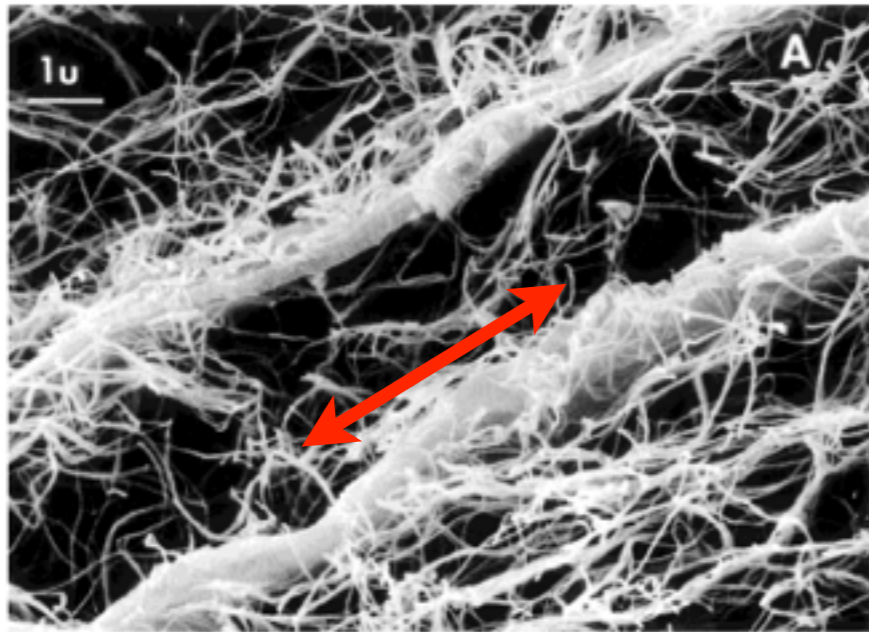
Van der Schaft et al. Tiss Eng A, 2011

Fibroblasts on collagen

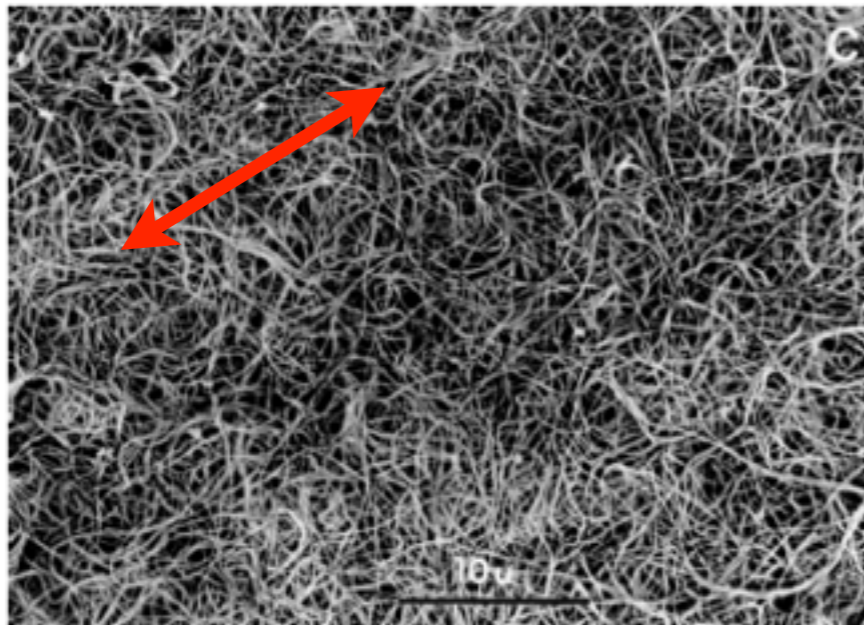


Eastwood et al. 1998

Fibroblasts align to stretch, but collagen fibers do not



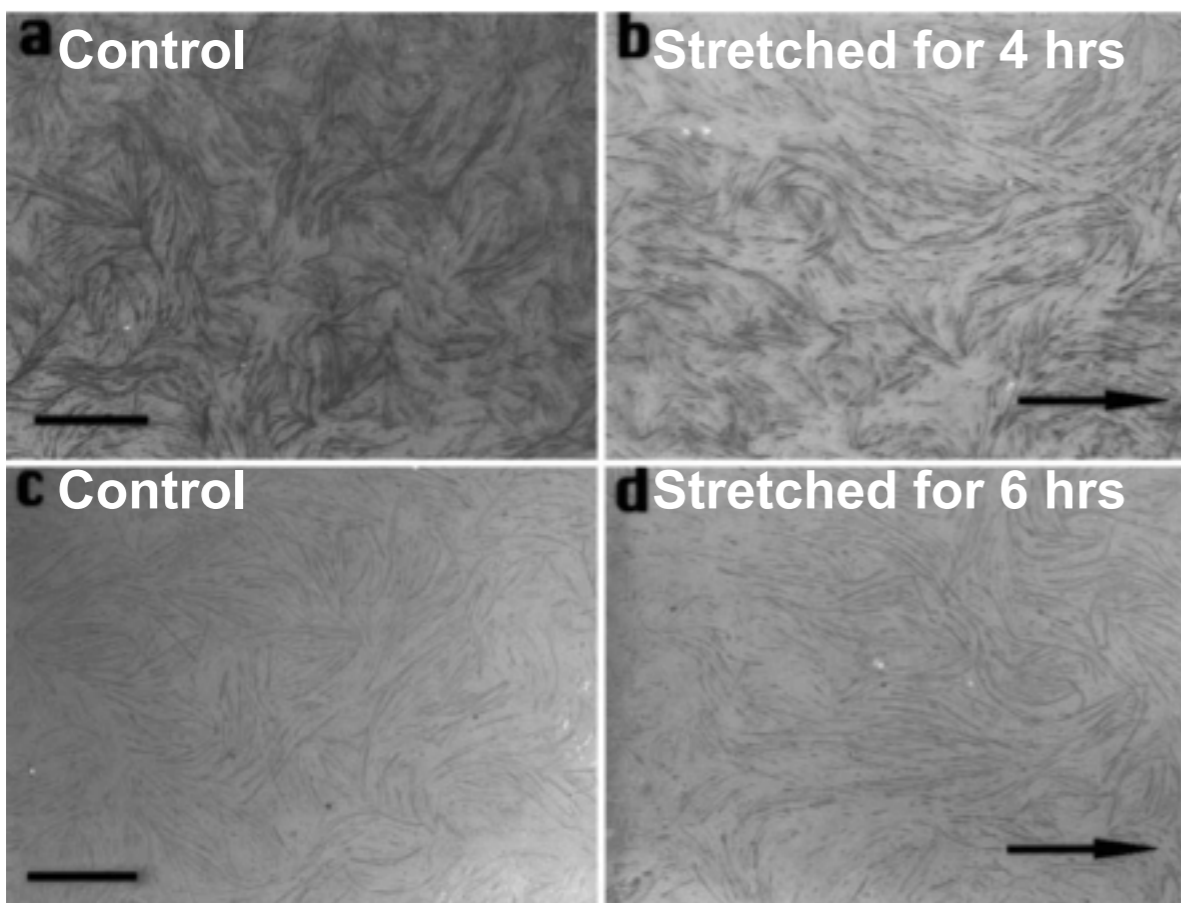
Cell-free matrix stretched overnight



Eastwood et al.
Cell Motil. Cytoskel. 1998

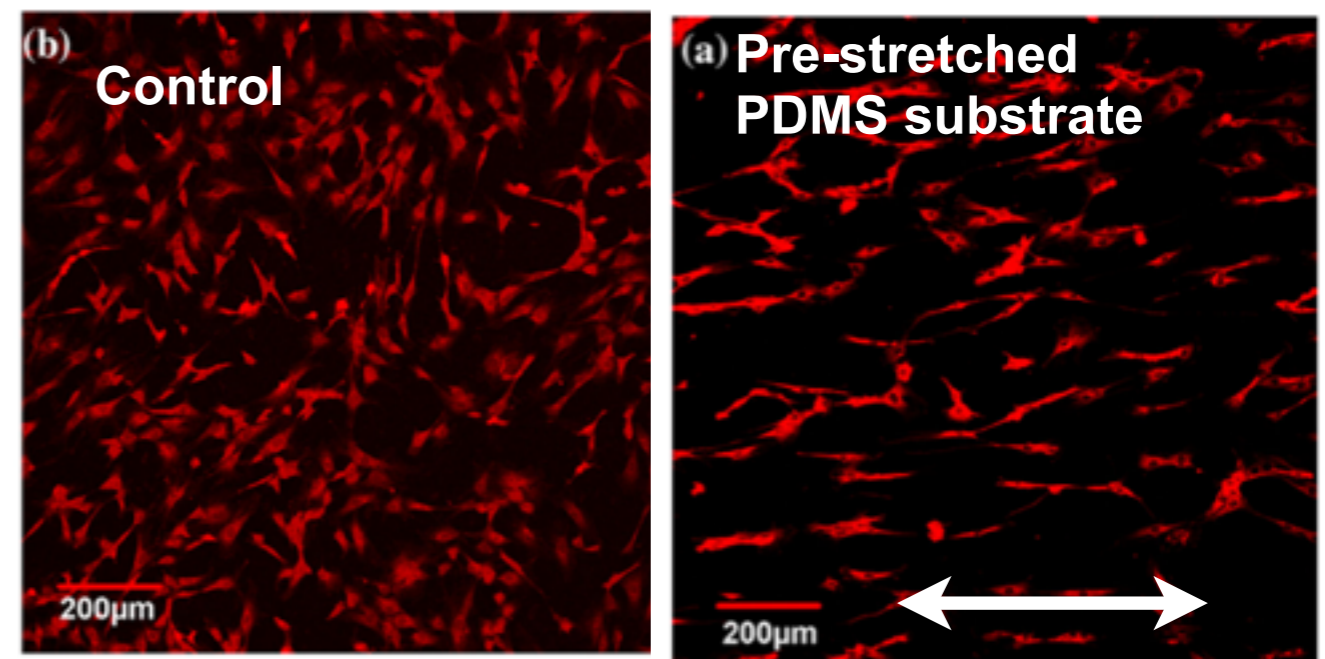
Cells also align to static stretch on **non-fibrous** substrates

Muscle fibers (mouse - Matrigel)



Collinsworth et al. Cell Tiss. Res. 2000

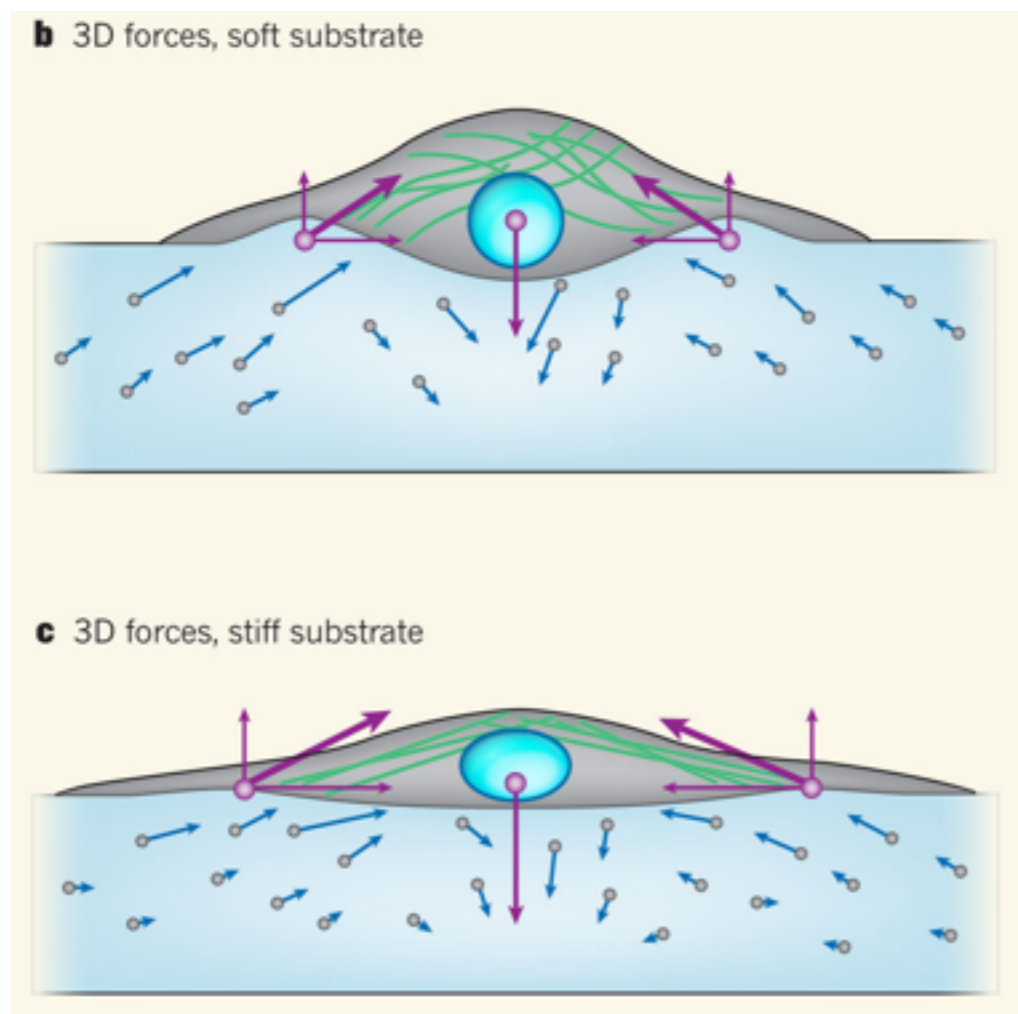
Mesenchymal stem cells (rat - PDMS)



Liu et al. Cell Mol Bioeng 2014

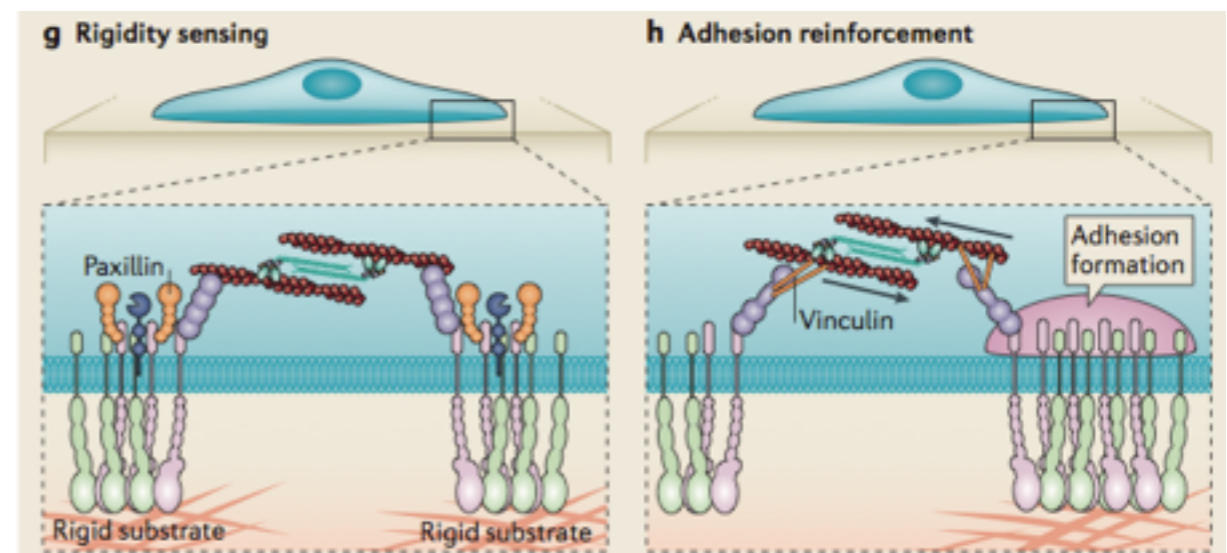
Hypothesis: 'active cell sensing'

- (1) Cells pull on matrix
- (2) matrix strain-stiffens



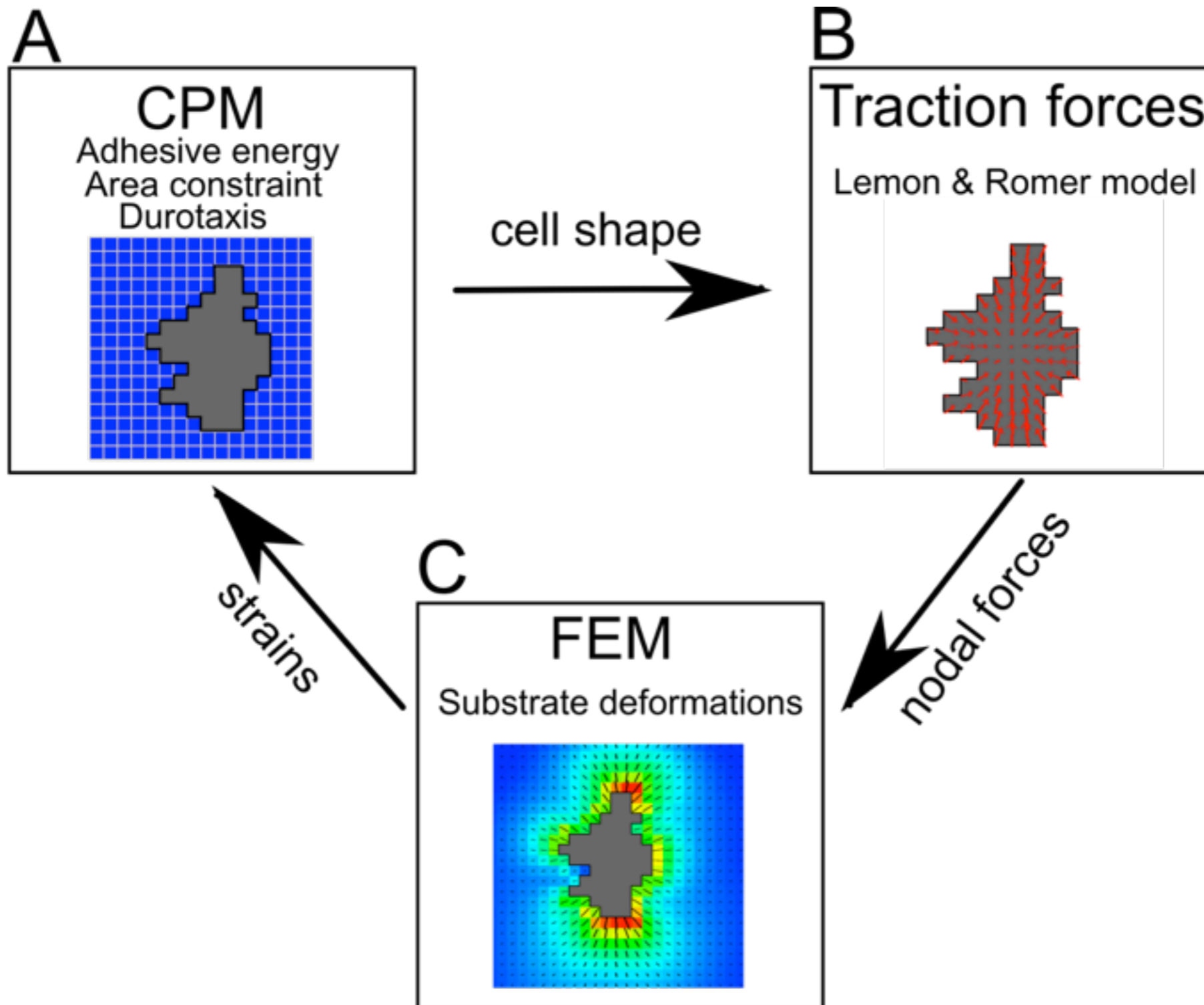
Hersen & Ladoux, Nature (2011)

- (3) Increased tension stabilizes focal adhesions to matrix on strained matrixes



Iskratsch et al. Nat. Rev. Mol. Cell. Biol. (2014)

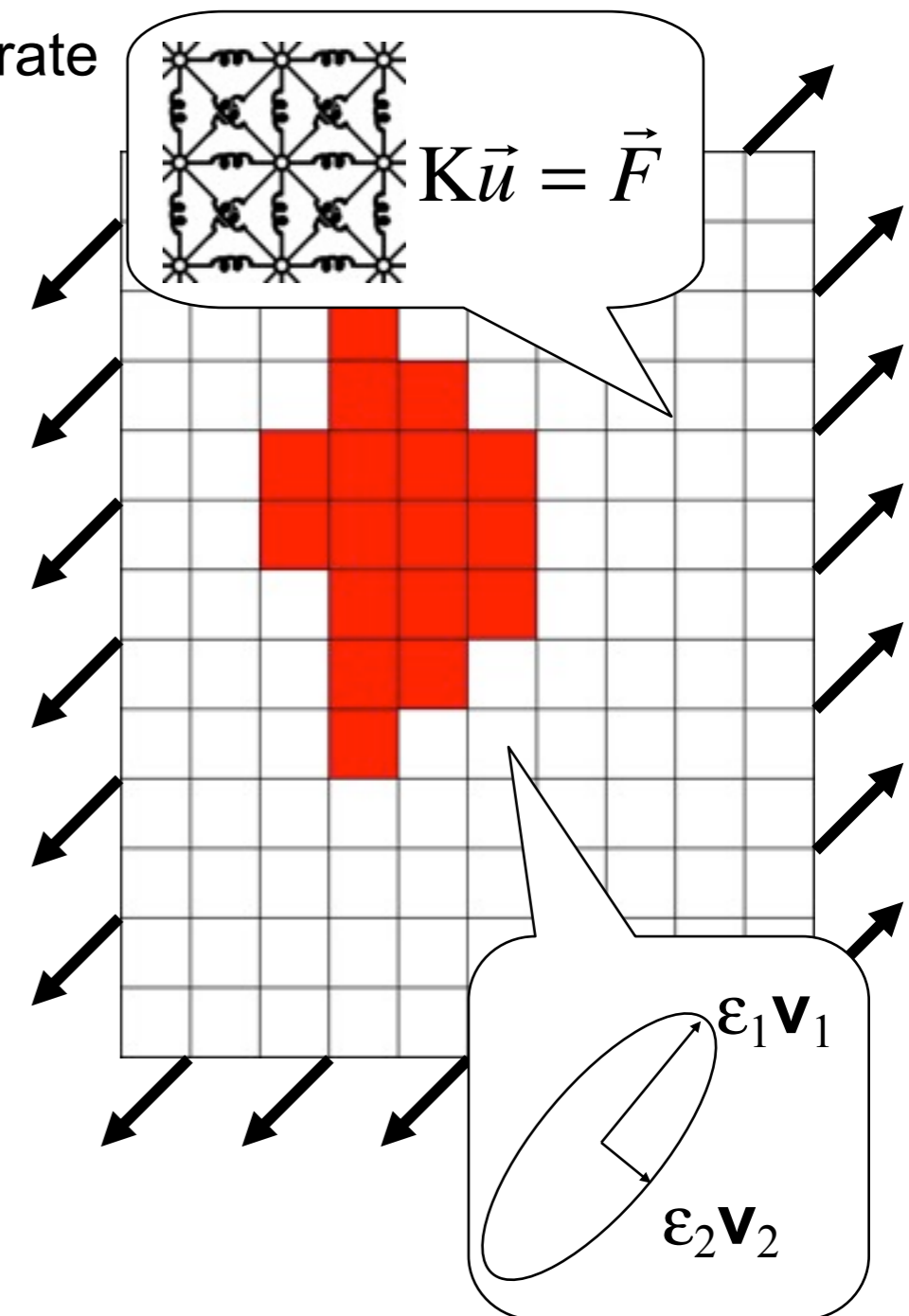
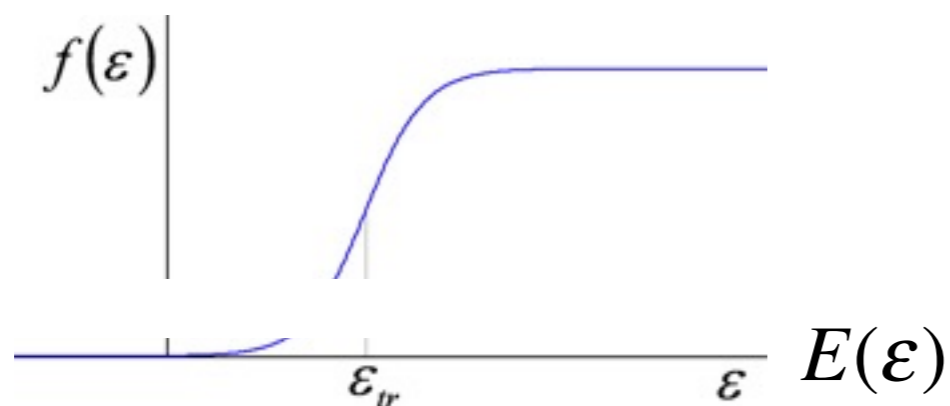
Mechanical cell-matrix feedback



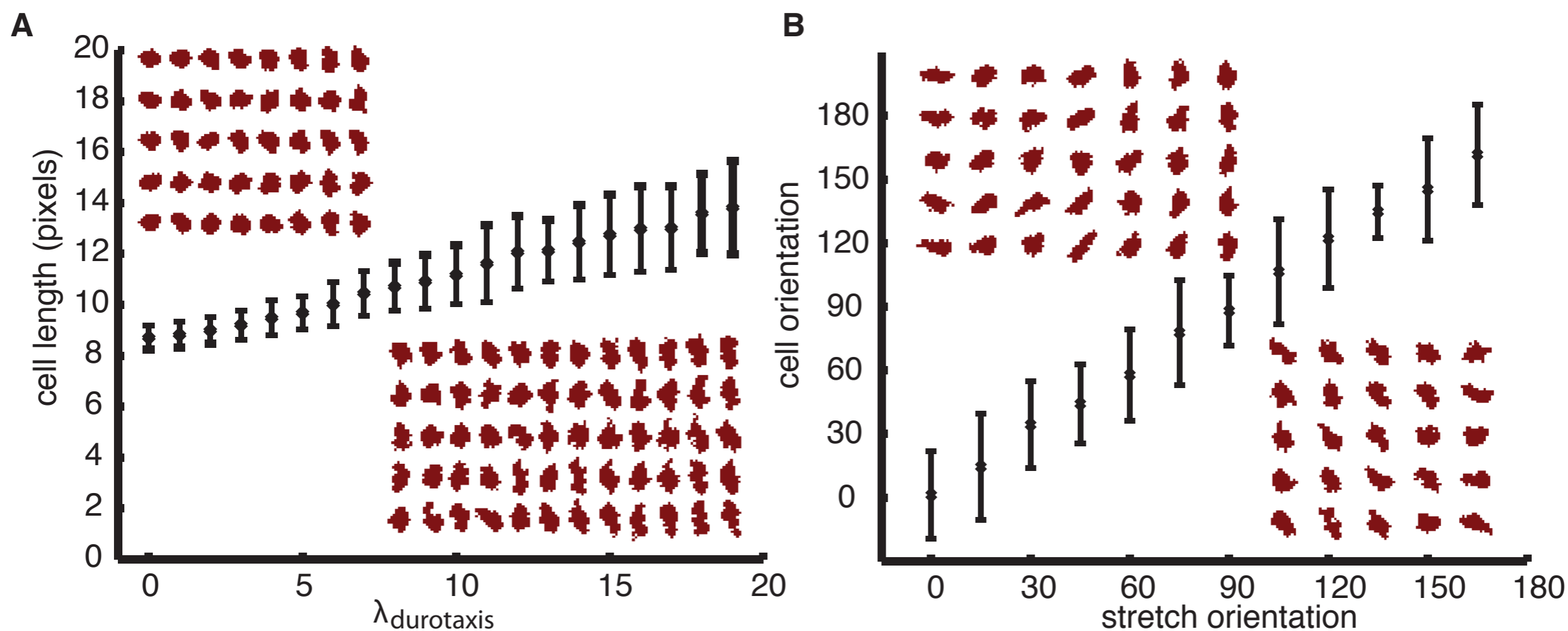
Modeling cell response to stretch using Cellular Potts Model

- ECM: Finite-element model (FE) of compliant substrate
 - Linear elastic assumptions
 - FE-calculations yield local principal strains
 - (magnitudes ε_1 and ε_2 , along \vec{v}_1 and \vec{v}_2)
 - Approximate strain stiffening:
 - Perceived ECM stiffness: $E(\varepsilon_1)$
 - Mimic focal adhesion maturation under strain:

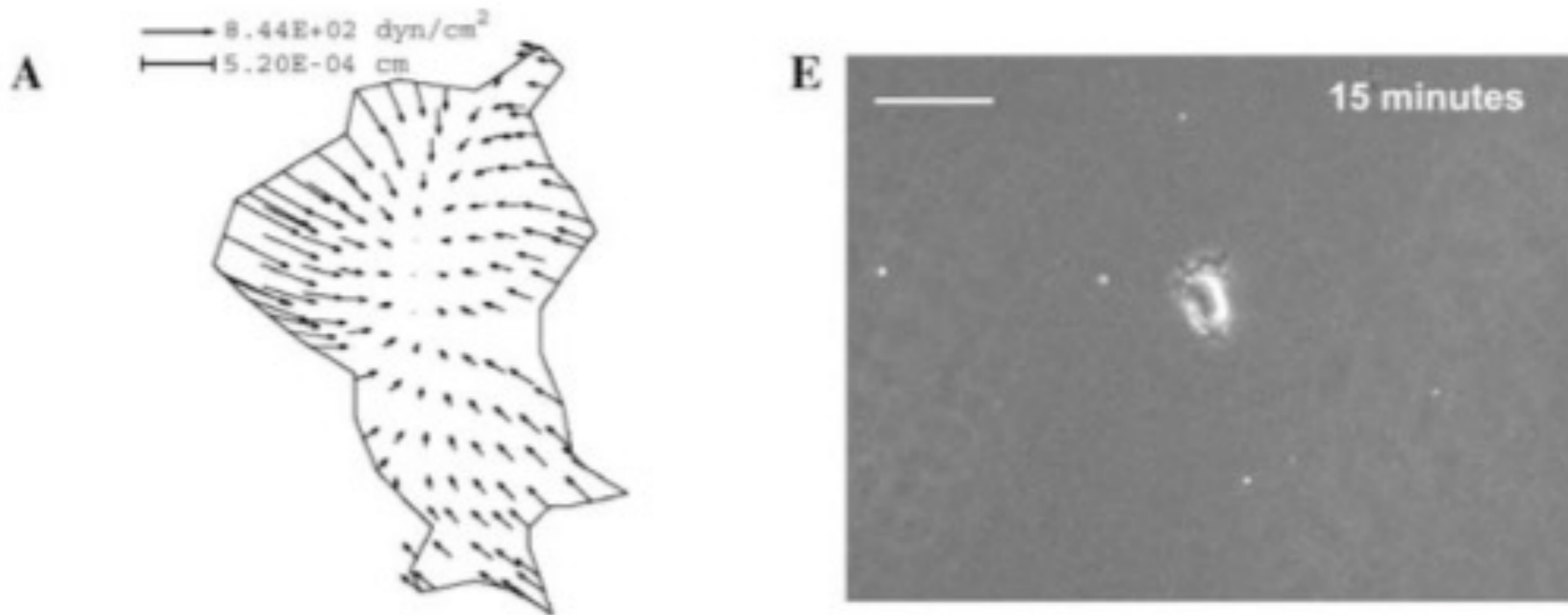
$$\Delta H_{\text{mech}} = -g(\vec{x}, \vec{x}') \lambda_{\text{durotaxis}} (f(E(\varepsilon_1))(\vec{v}_1 \cdot \vec{v}_m)^2 + f(E(\varepsilon_2))(\vec{v}_2 \cdot \vec{v}_m)^2)$$



Response of single cells to external, static strain



Cells pull on substrate

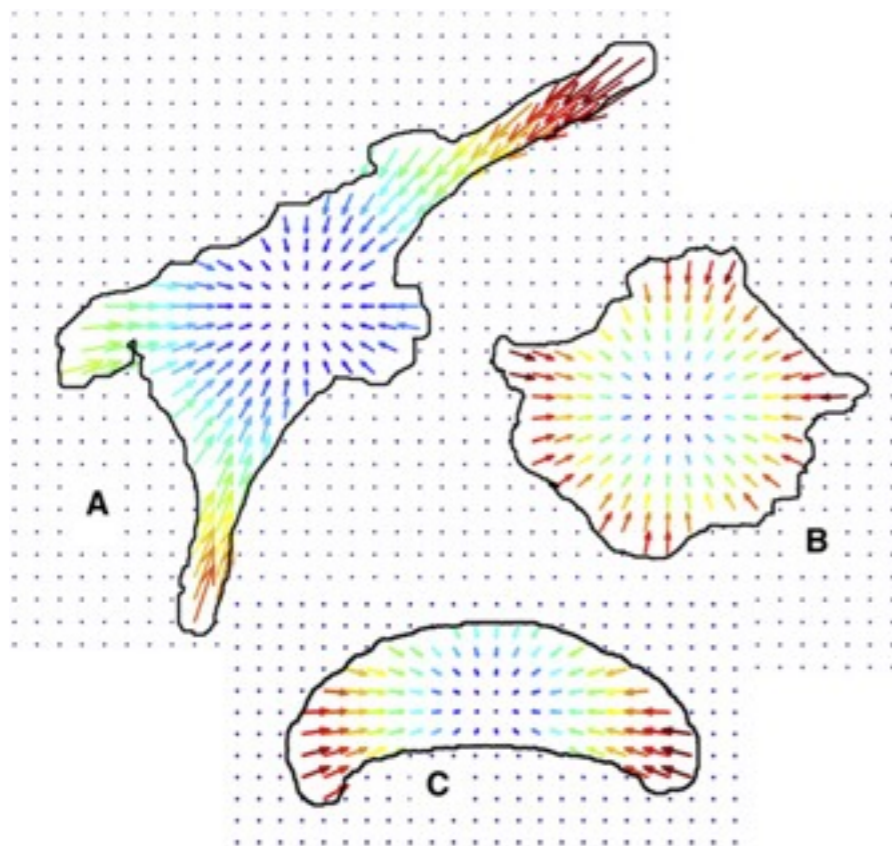


Reinhart-King et al. Biophys. J. 2005

Van Oers, Rens, et al. *PLoS Comp Biol.* 2014

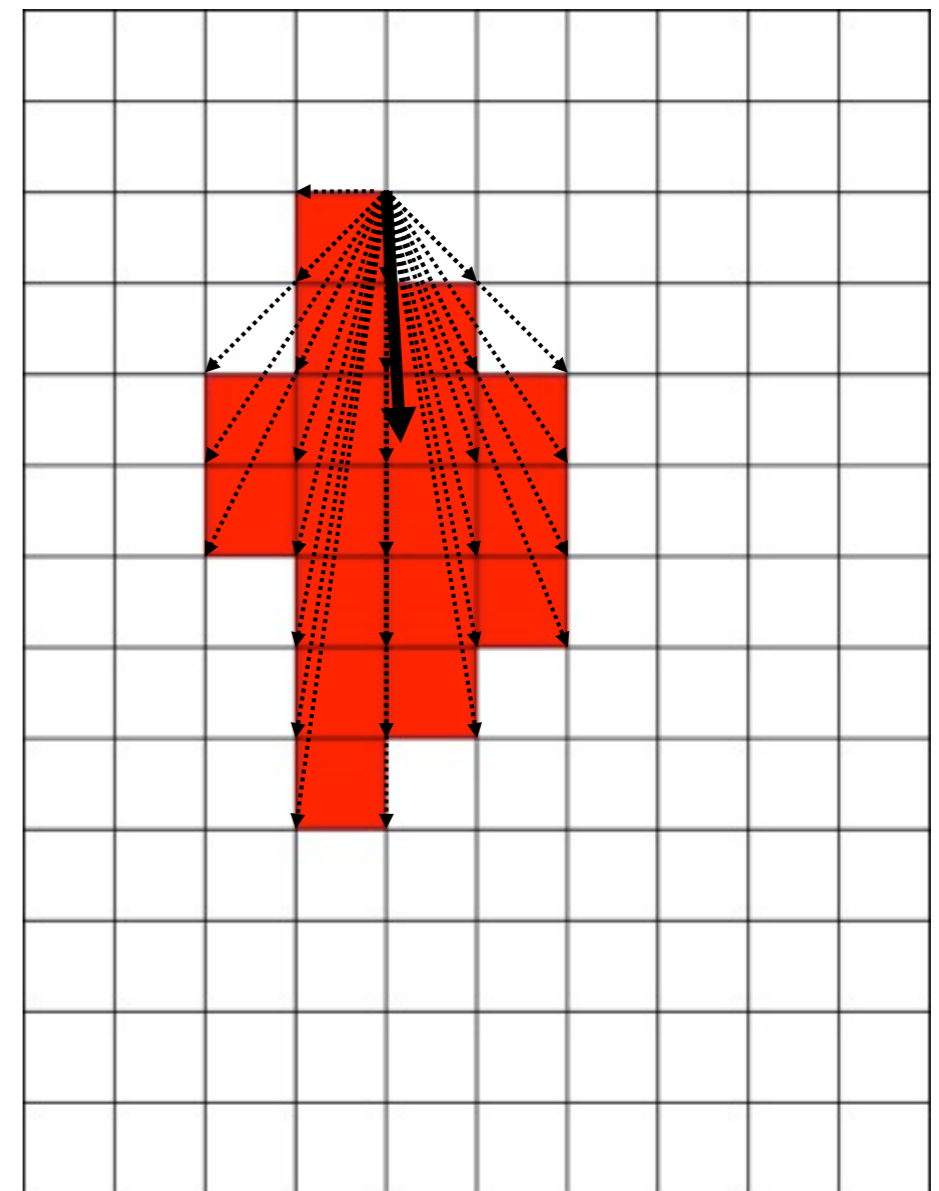
Cells pull on substrate

- Lemmon & Romer (*Biophys. J.* 2010):
 - Cells “acts as single cohesive unit”
 - Force between any two points in cell proportional to distance between them
 - Zero traction in cell centroid



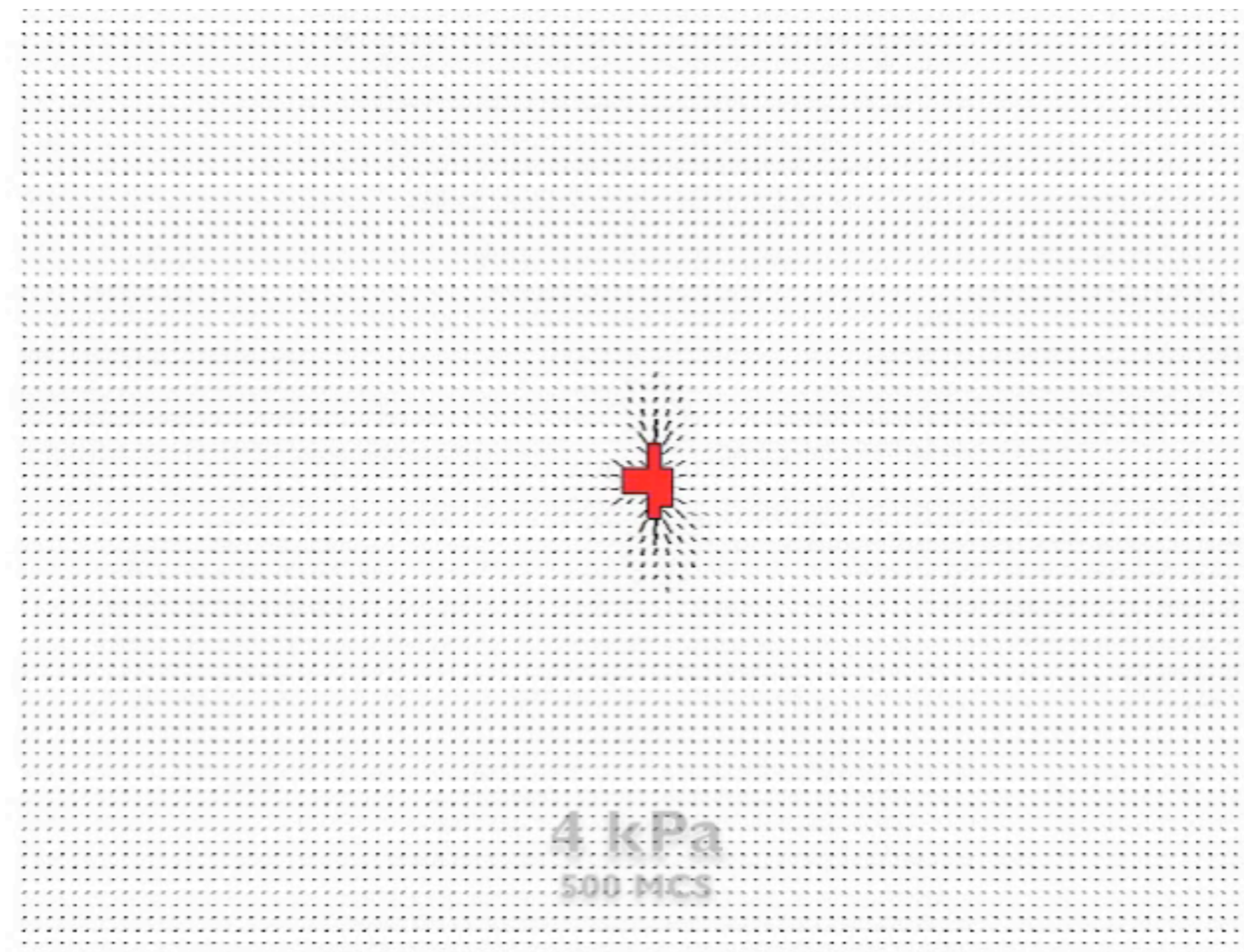
$$\mathbf{F}_i = \mu \sum_j \mathbf{d}_{i,j}$$

Implementation in CPM:



Van Oers, Rens, et al. *PLoS Comp Biol.* 2014

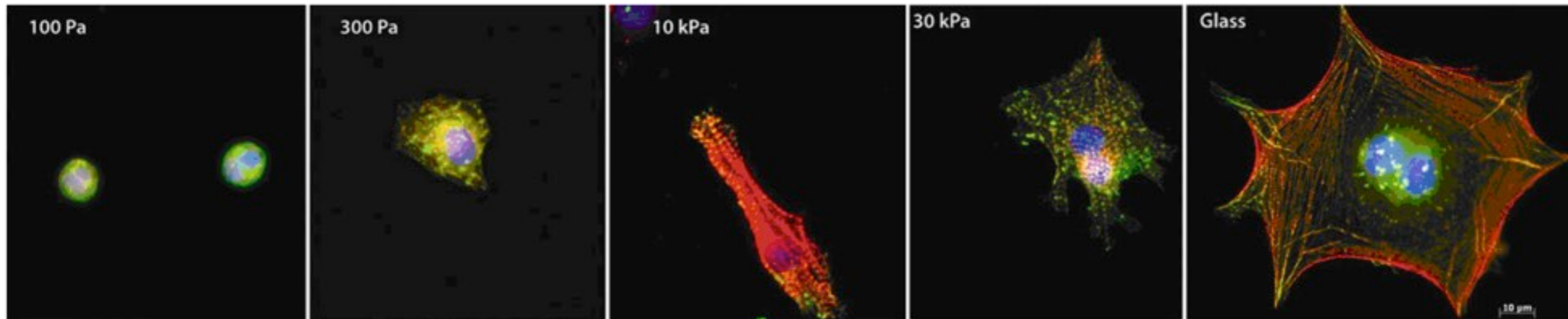
Feedback between cell traction and strain response



Behavior of single cells

Feedback between cell-induced strain and cell responses

Cardiomyocytes (it works about the same for ECs...):



Winer et al. , in: Wagoner et al. (eds.), 2011

SOFT SUBSTRATE

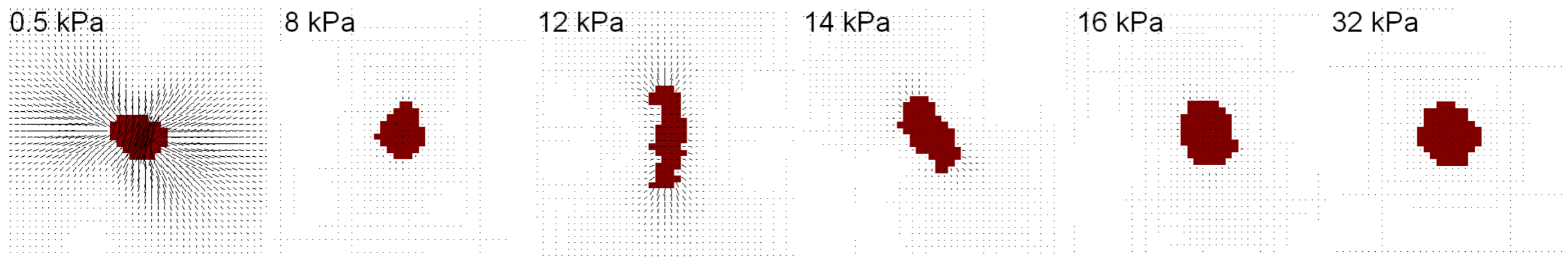
- stretch all around
- contraction

INTERMEDIATE SUBS.

- stretch along long axis
- elongation

STIFF SUBSTRATE

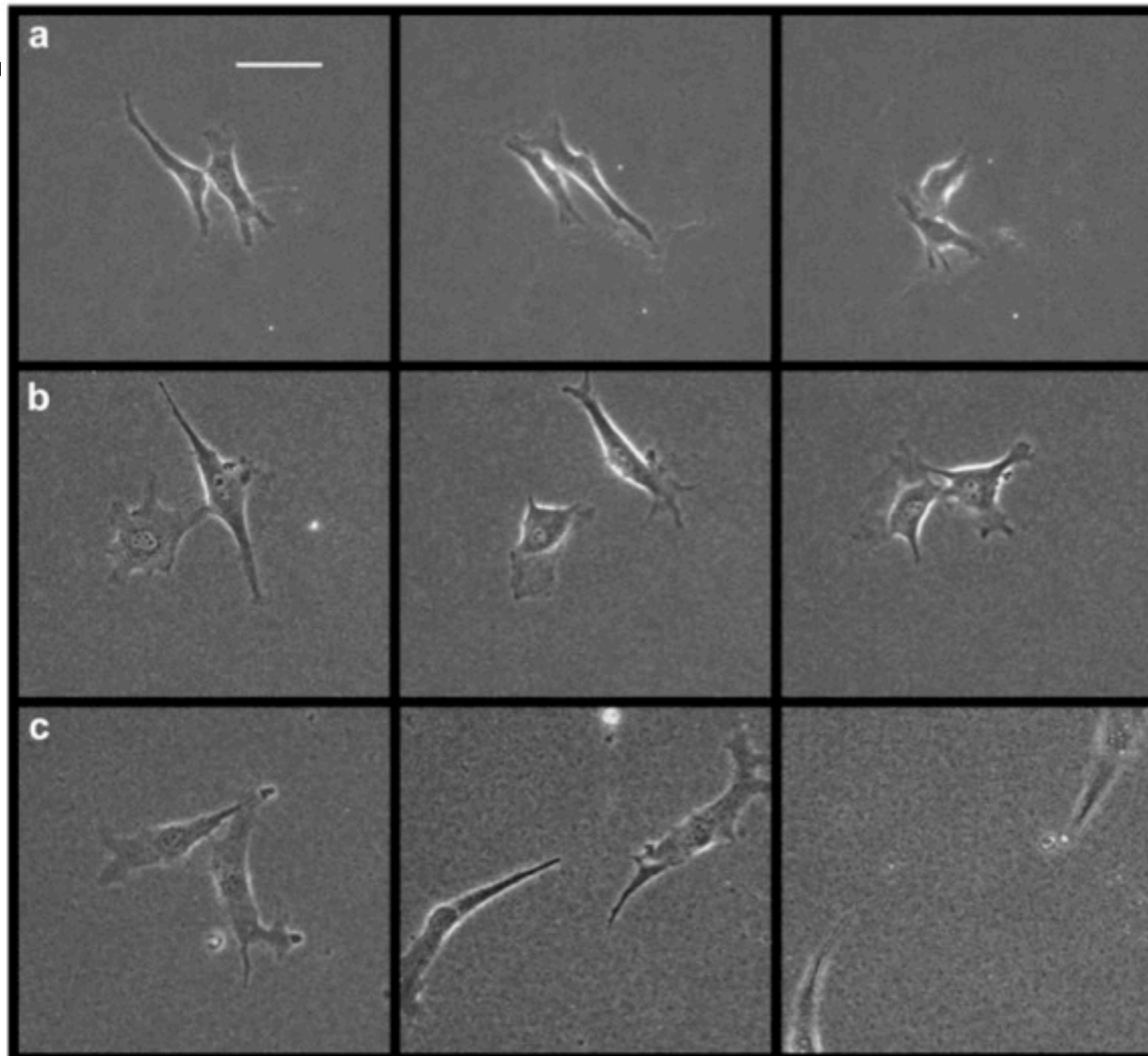
- little stretch
- spreading



Van Oers, Rens, et al. *PLoS Comp Biol.* 2014

Cell-cell interactions

(Reinhart-King et al. 2008)



Soft matrix (500 Pa)

Cells touch and remain in contact

Stiffer matrix (5.5 kPa)

Cells touch, loose contact, touch again

Stiff matrix (33 kPa)

Cells touch and walk away

Mechanical cell-cell communication

Video S2 - cell pairs

Van Oers, Rens, LaValley, Reinhart-King, and Merks
PLoS Comp Biol 2014



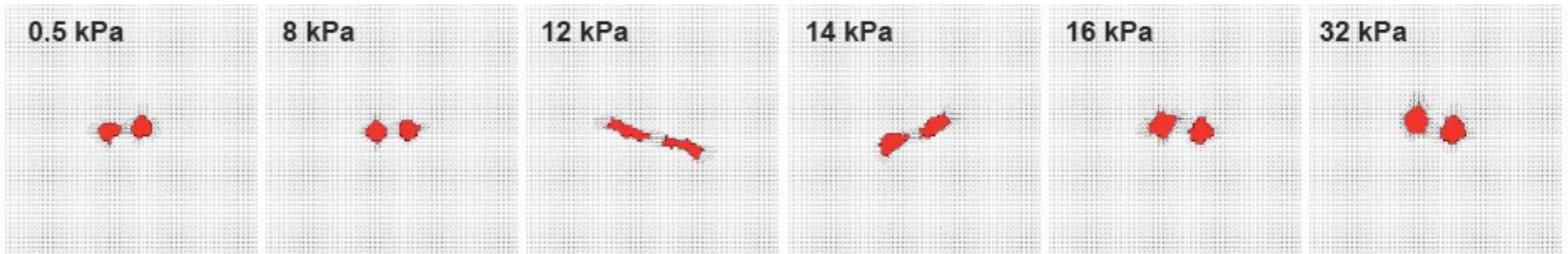
cf. Bischofs and Schwarz
PNAS 2003

Van Oers, Rens, et al. *PLoS Comp Biol.* 2014

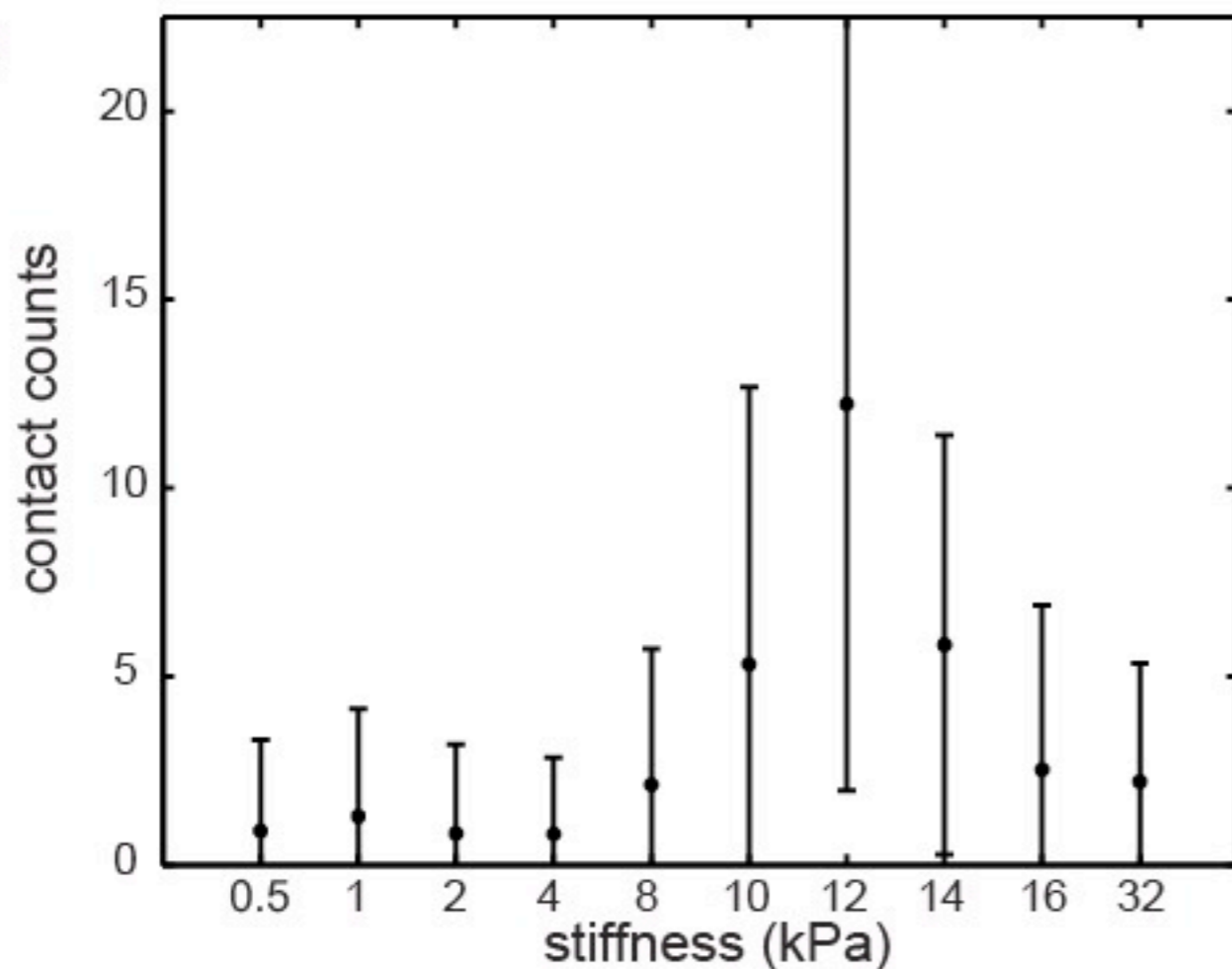
Mechanical cell-cell communication

Universiteit Leiden

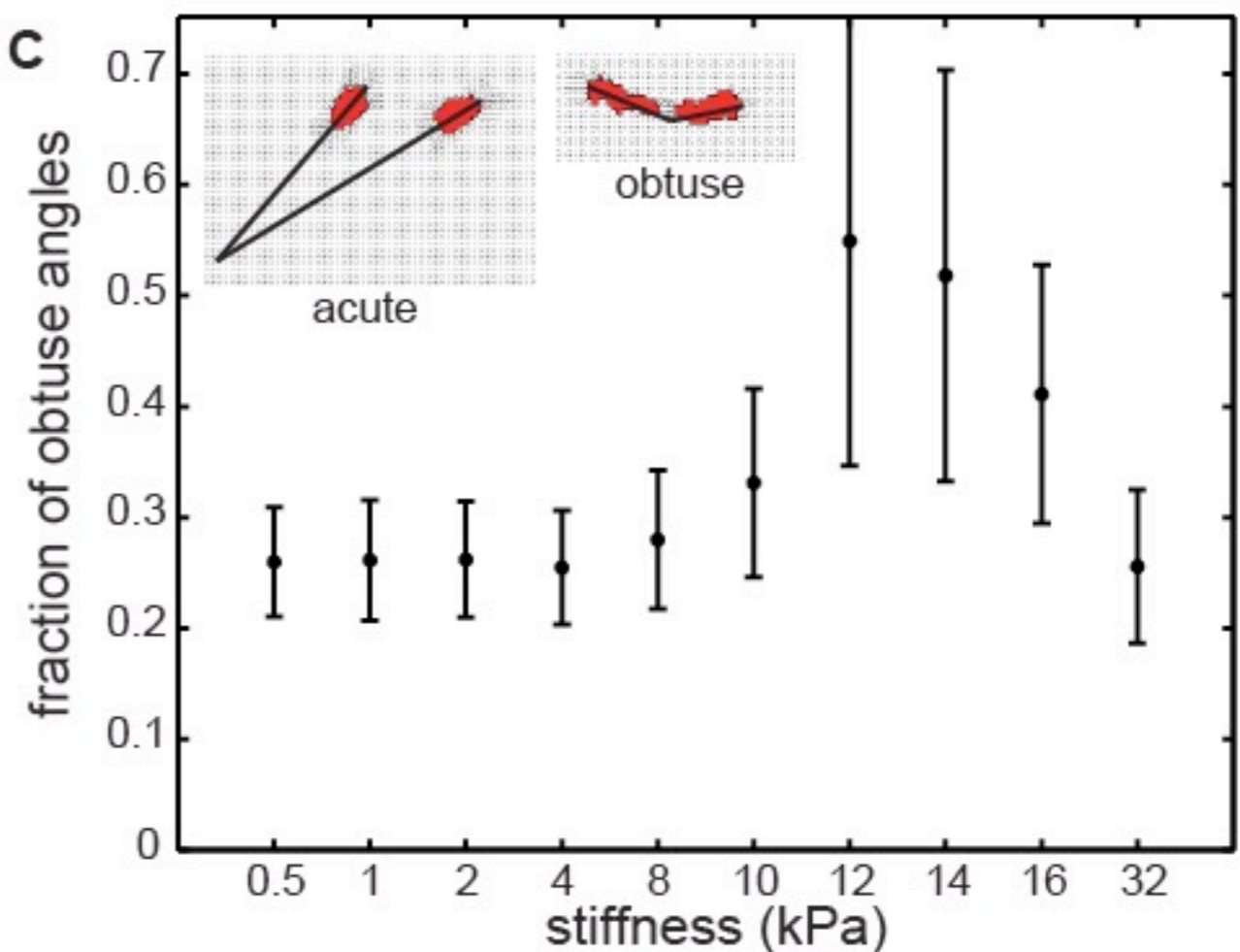
A



B



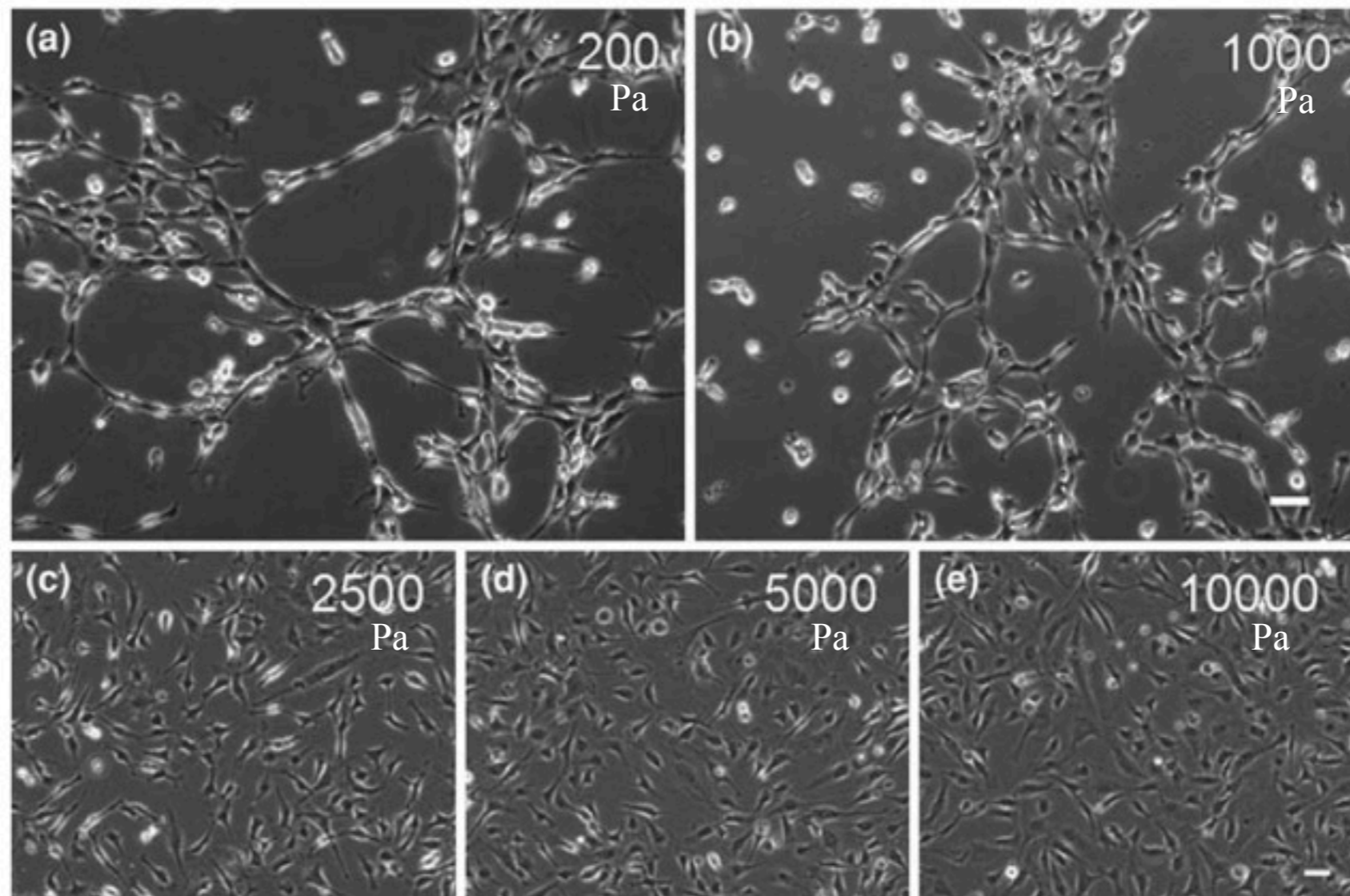
C

Van Oers, Rens, et al. *PLoS Comp Biol.* 2014

Collective cell behavior on unstretched matrix

Example: bovine aortic endothelial cells on
poly-acrylamide substrate (non-fibrous)

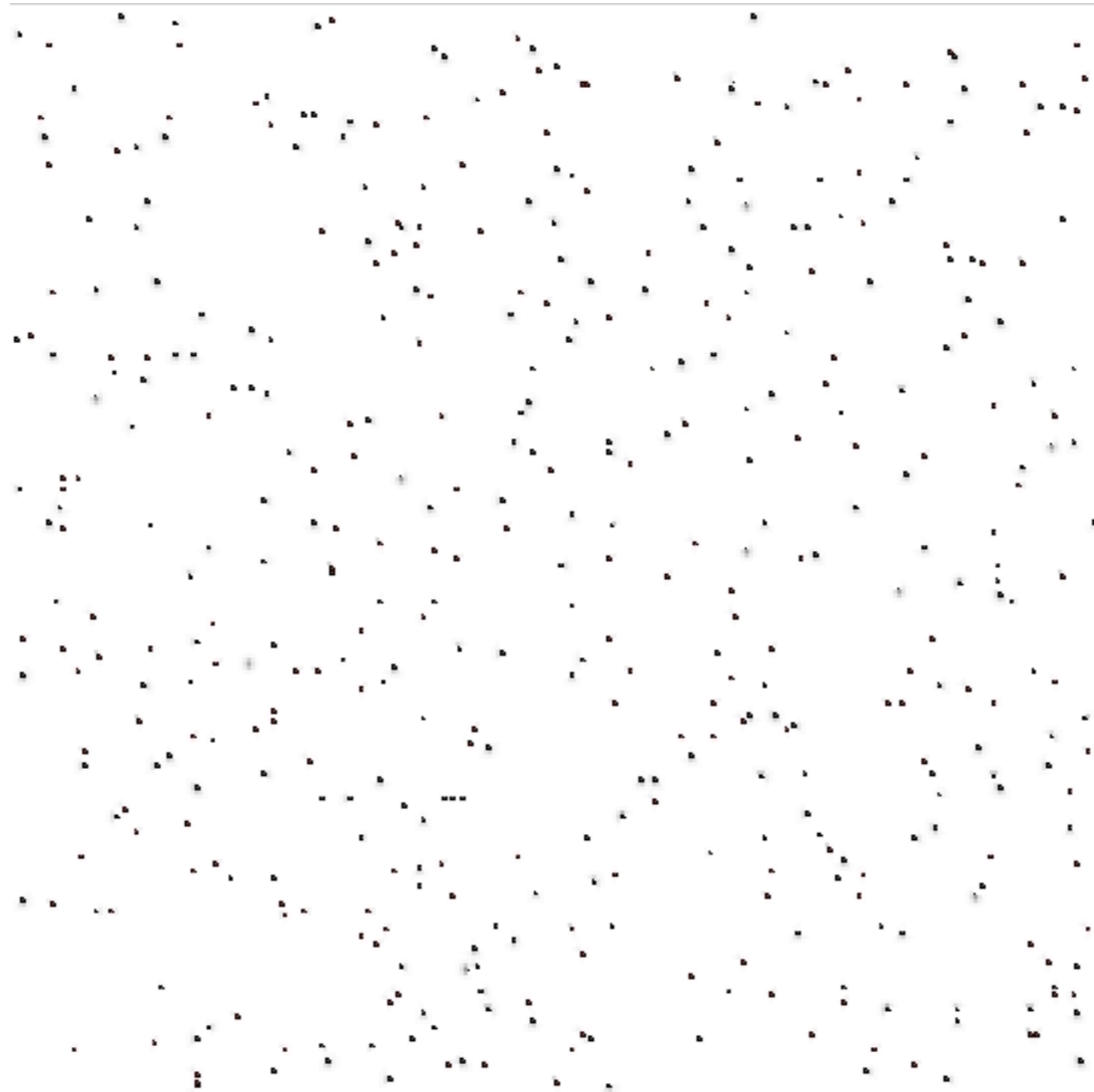
Soft matrix



Stiff matrix

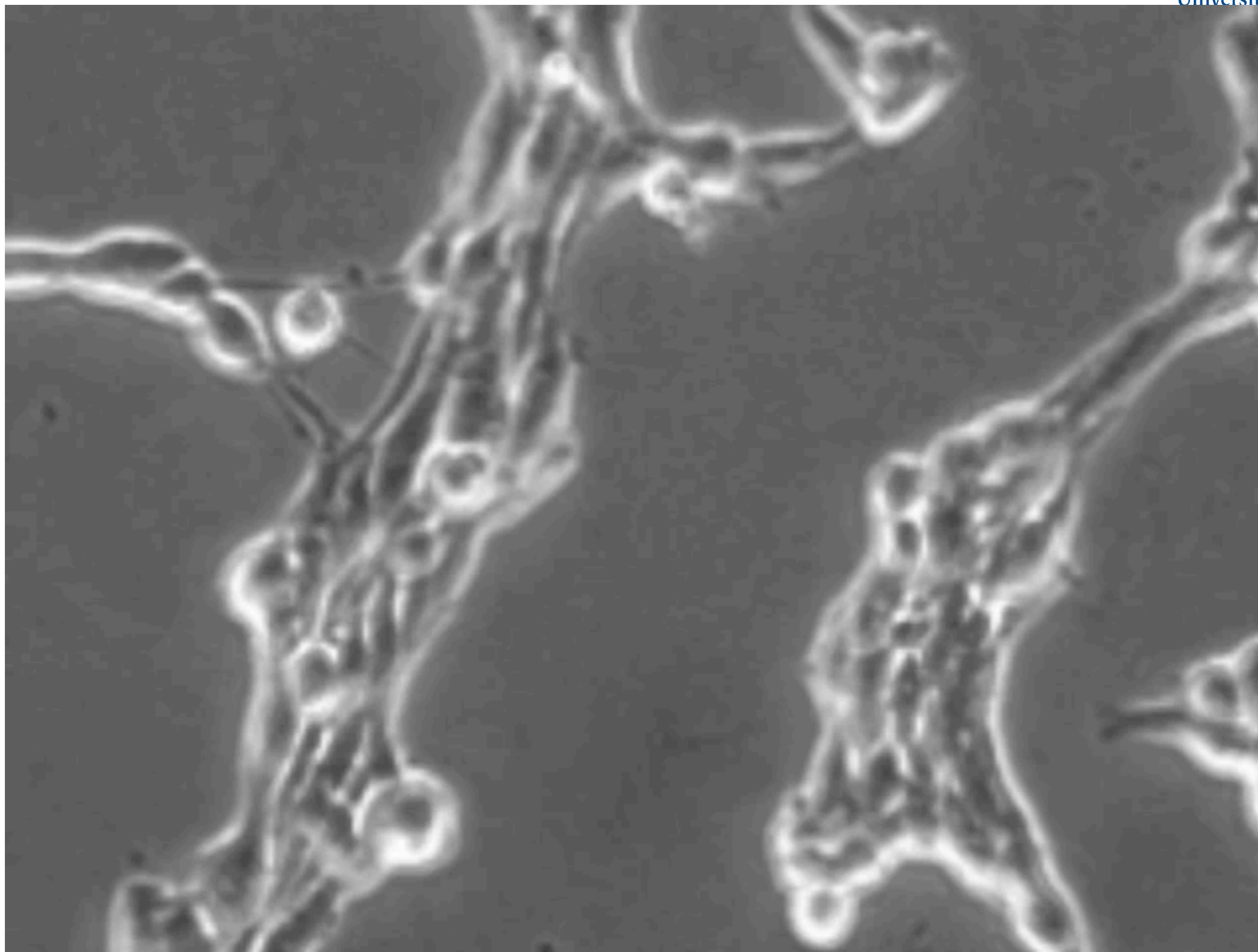
Califano and Reinhart-King, 2008

Resulting collective behavior

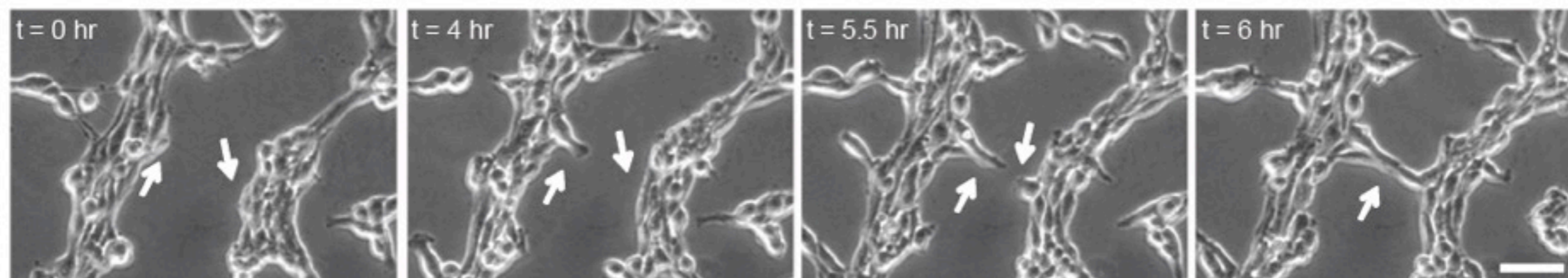
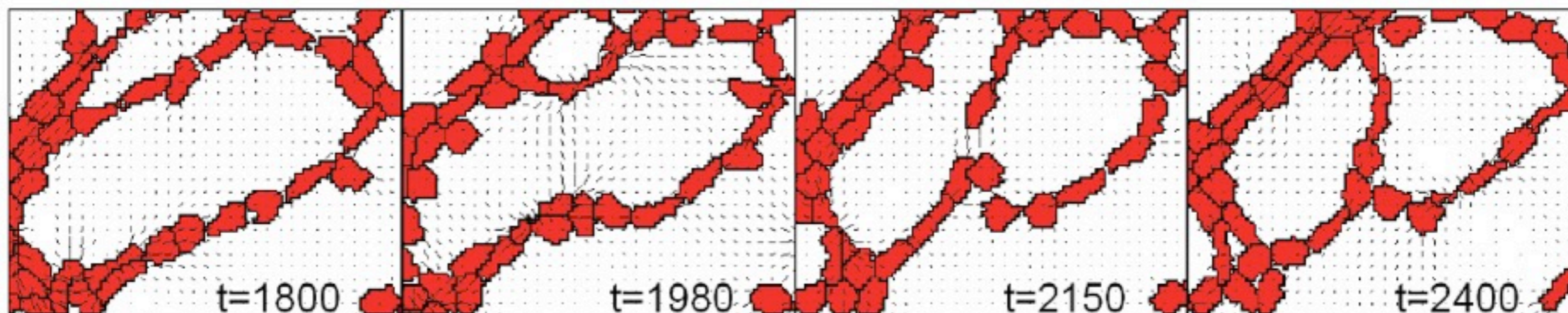


10 kPa

Van Oers, Rens, et al. *PLoS Comp Biol.* 2014

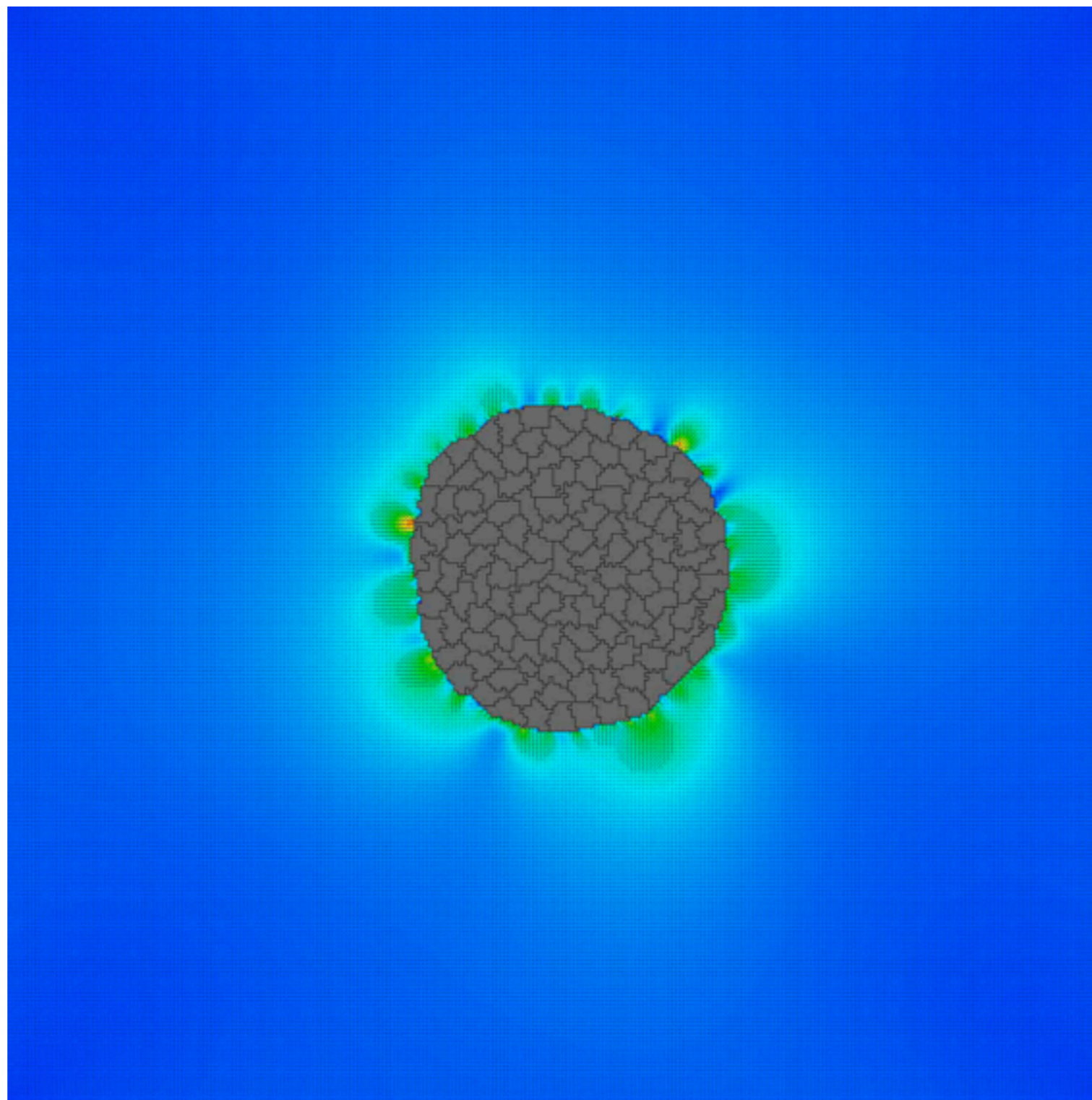


Bridging events



Danielle LaValley and
Cynthia Reinhart-King

Sprouting

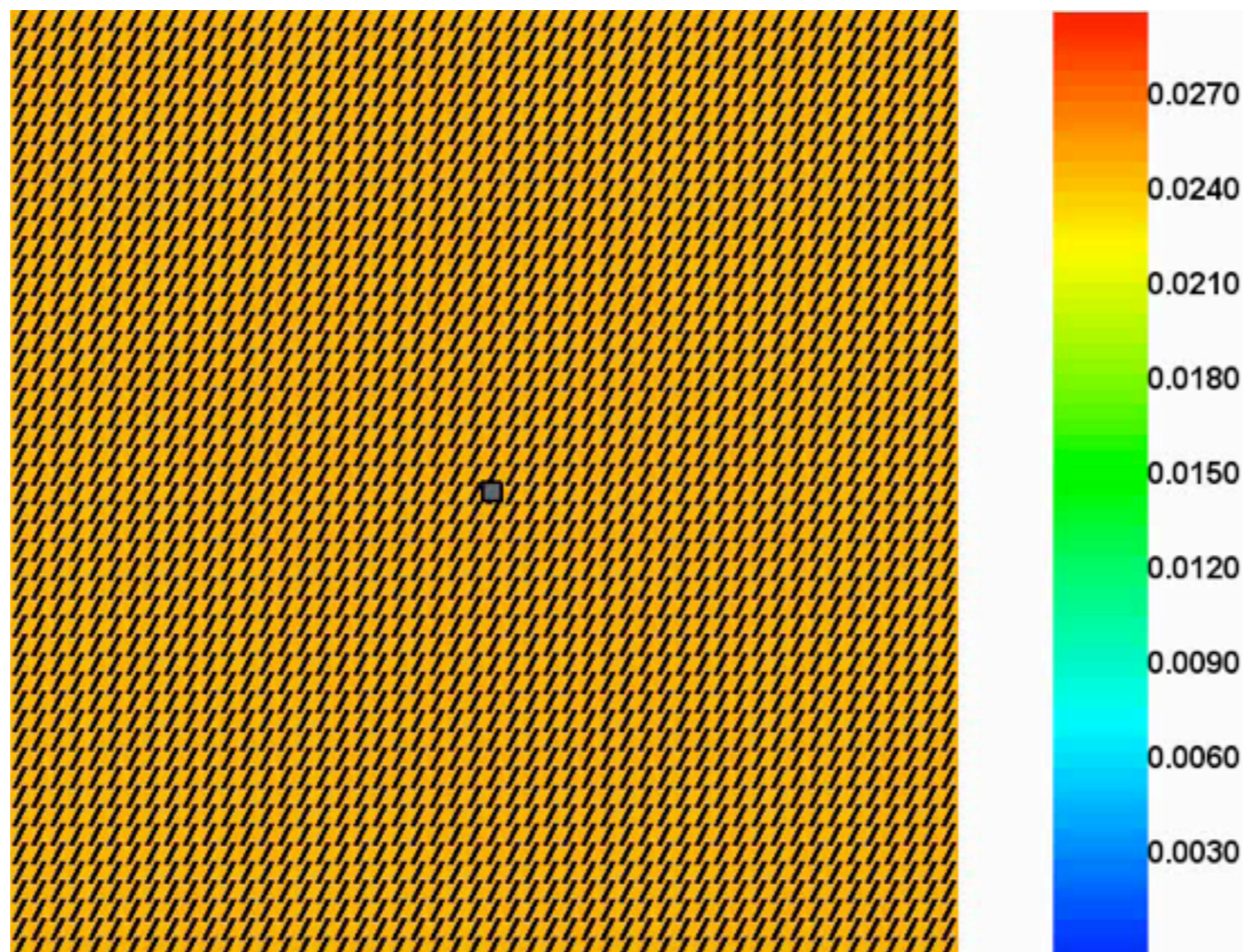


10 kPa

Van Oers, Rens, et al. *PLoS Comp Biol.* 2014

Cell contractility (“active sensing”) amplifies sensitivity to matrix strain

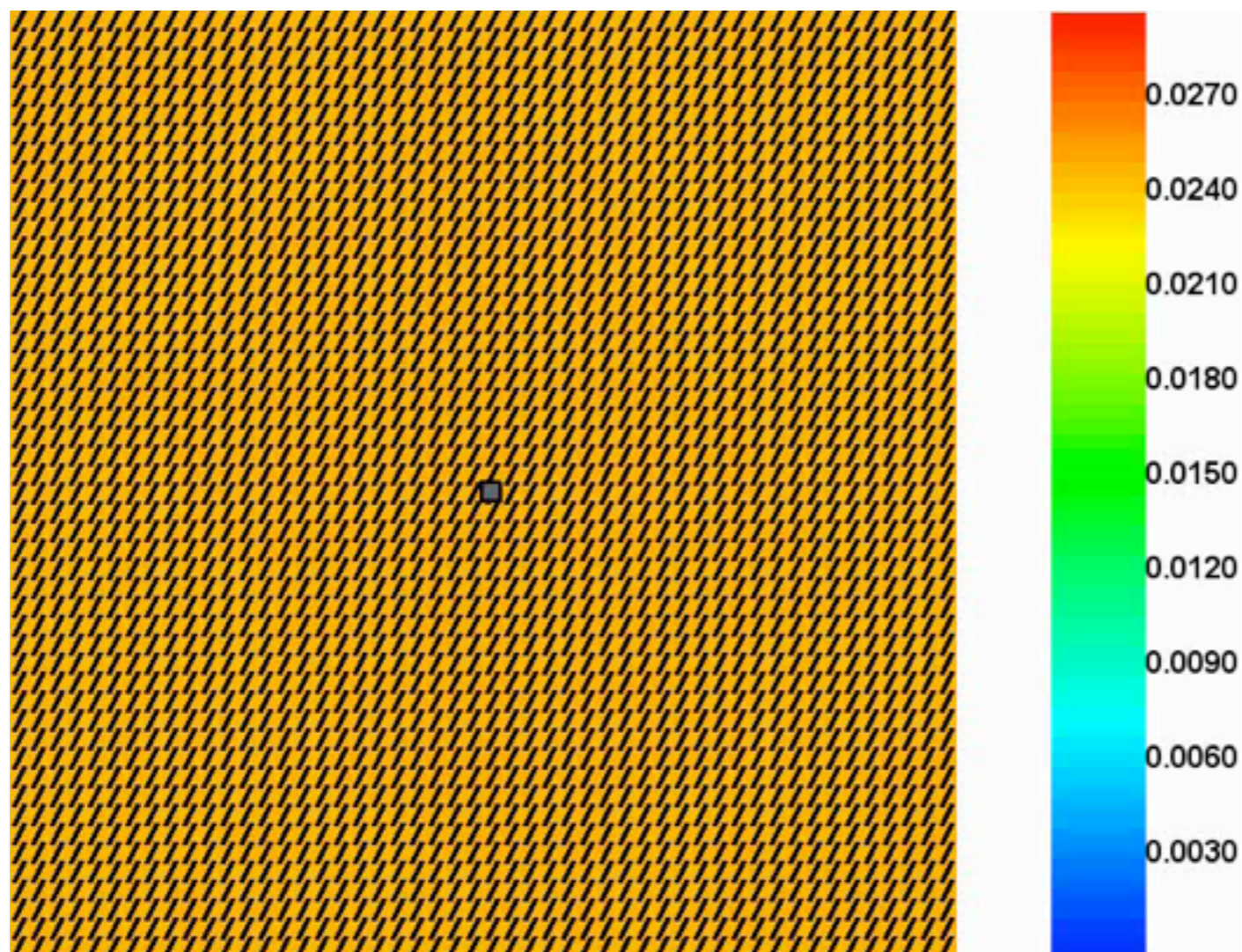
- Only external strain - 30 degrees, 0.025
- Cells do not exert forces on matrix



Rens & Merks
Biophys. J. 2017
arXiv:1605.03987

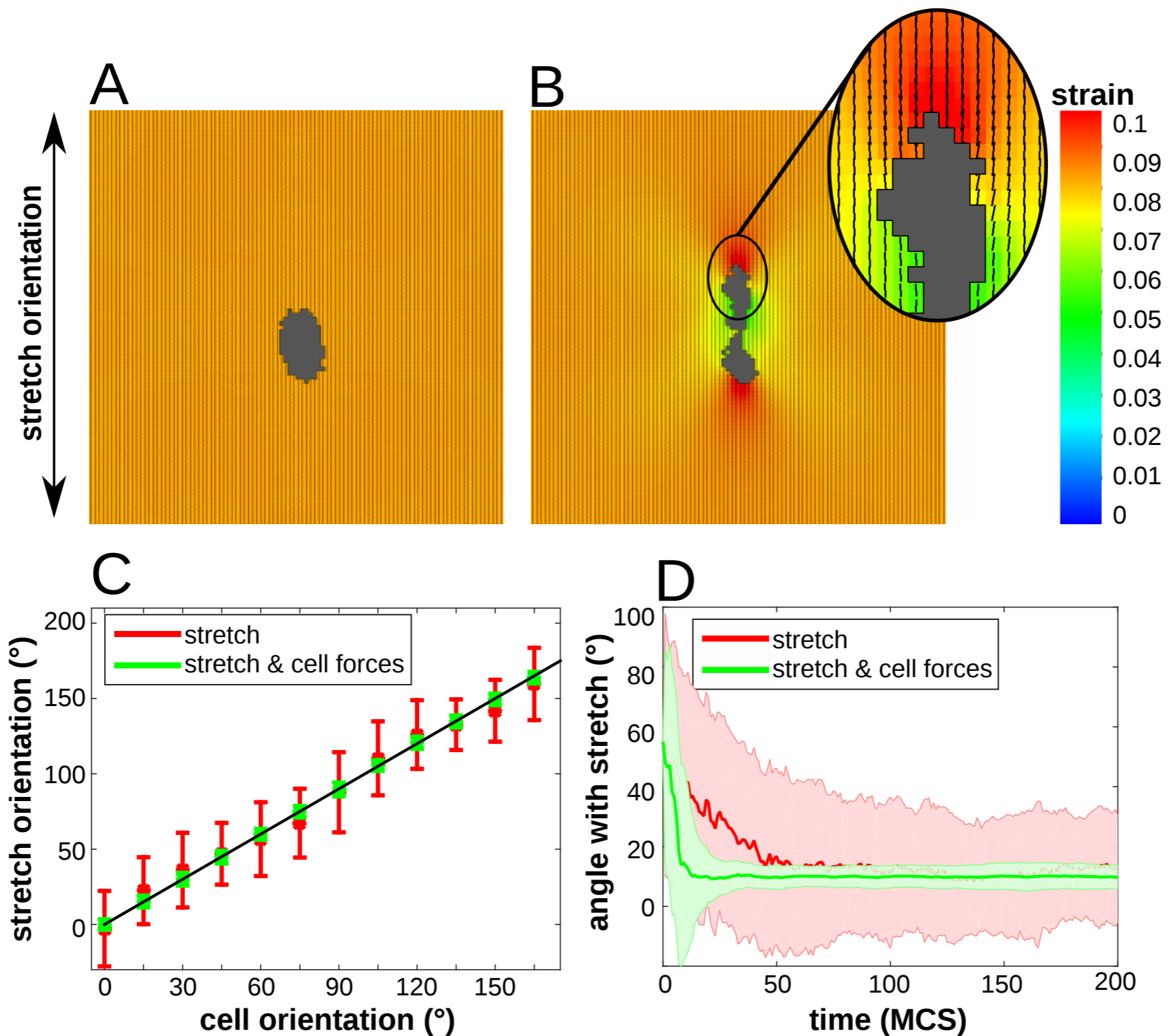
Cell contractility (“active sensing”) amplifies sensitivity to matrix strain

- External strain - 0.025, 30 degrees
- Cells **do** exert forces on matrix



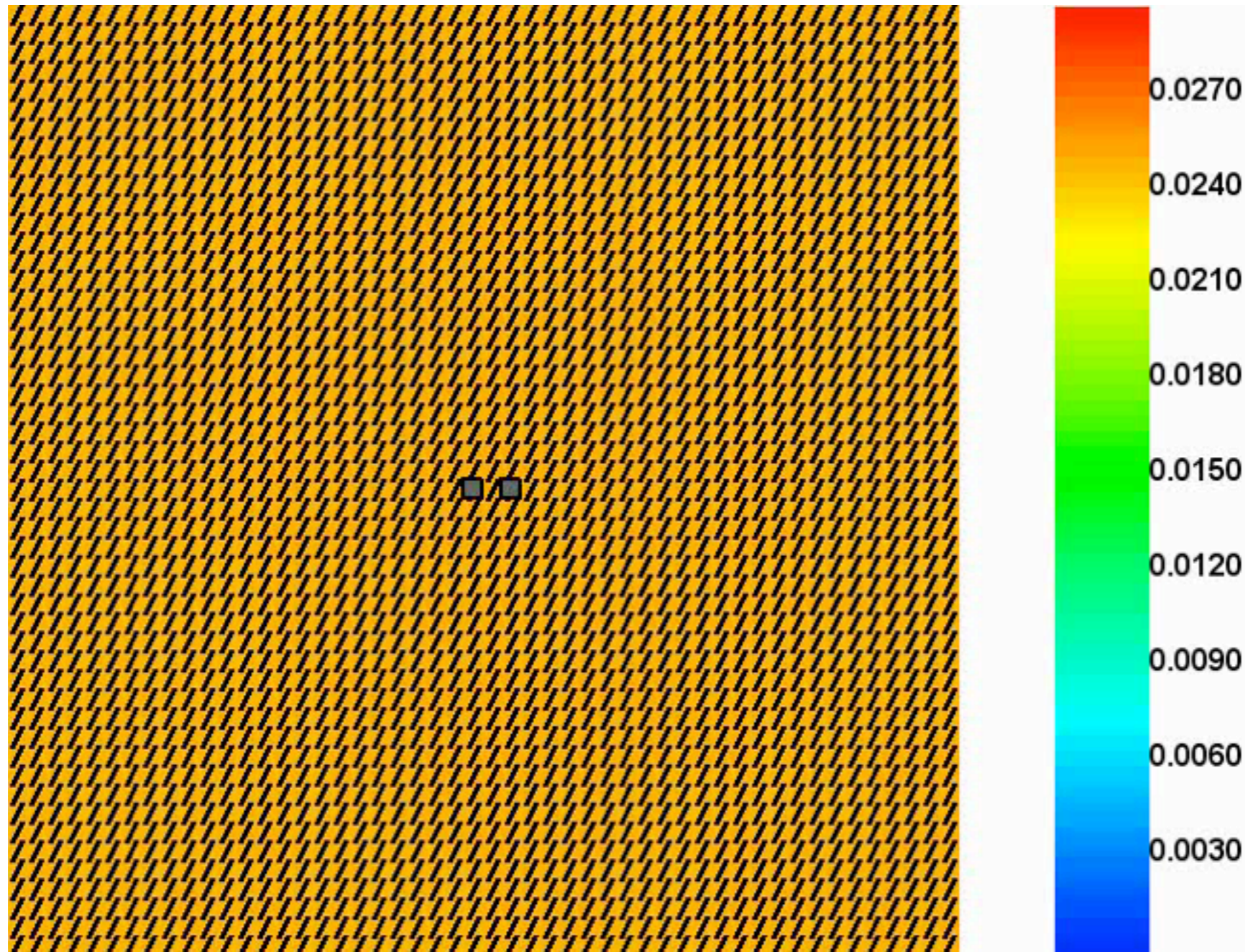
Rens & Merks, Biophys. J. 2017, arXiv:1605.03987

Contractility increases precision and speed of cell orientation



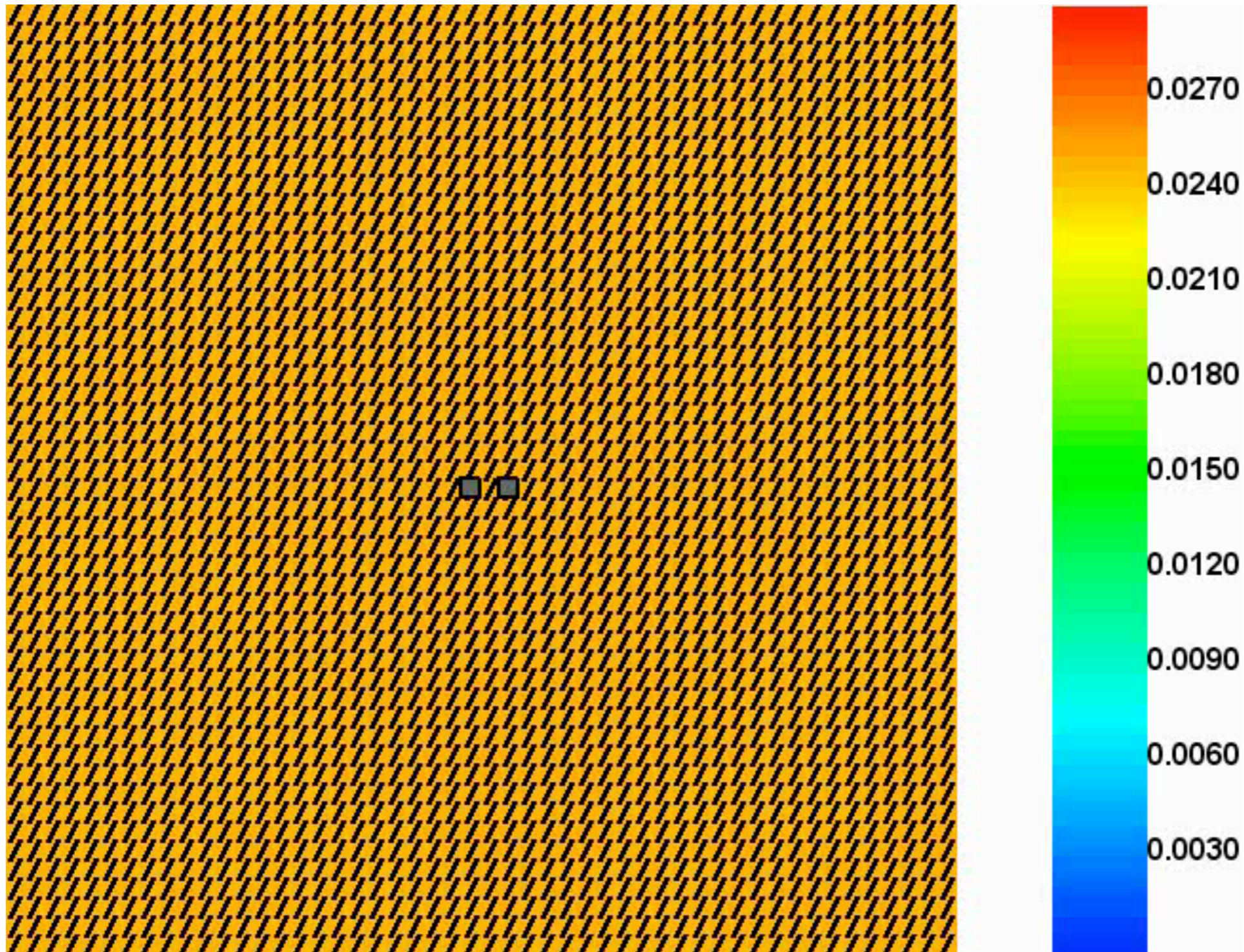
Rens & Merks, Biophys. J. 2017, arXiv:1605.03987

Two cells, no cellular traction



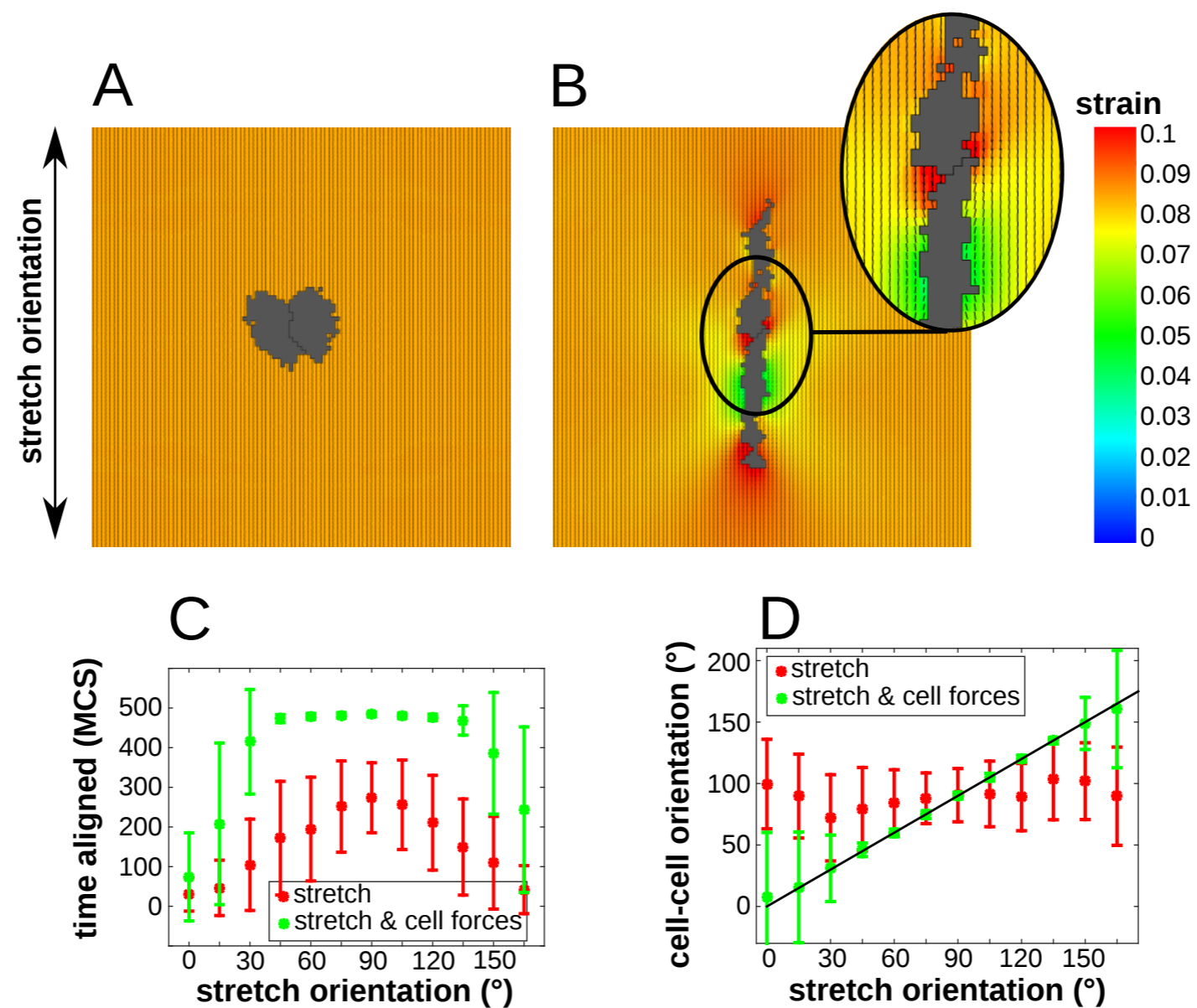
Rens & Merks, Biophys. J. 2017, arXiv:1605.03987

Two cells, cellular traction



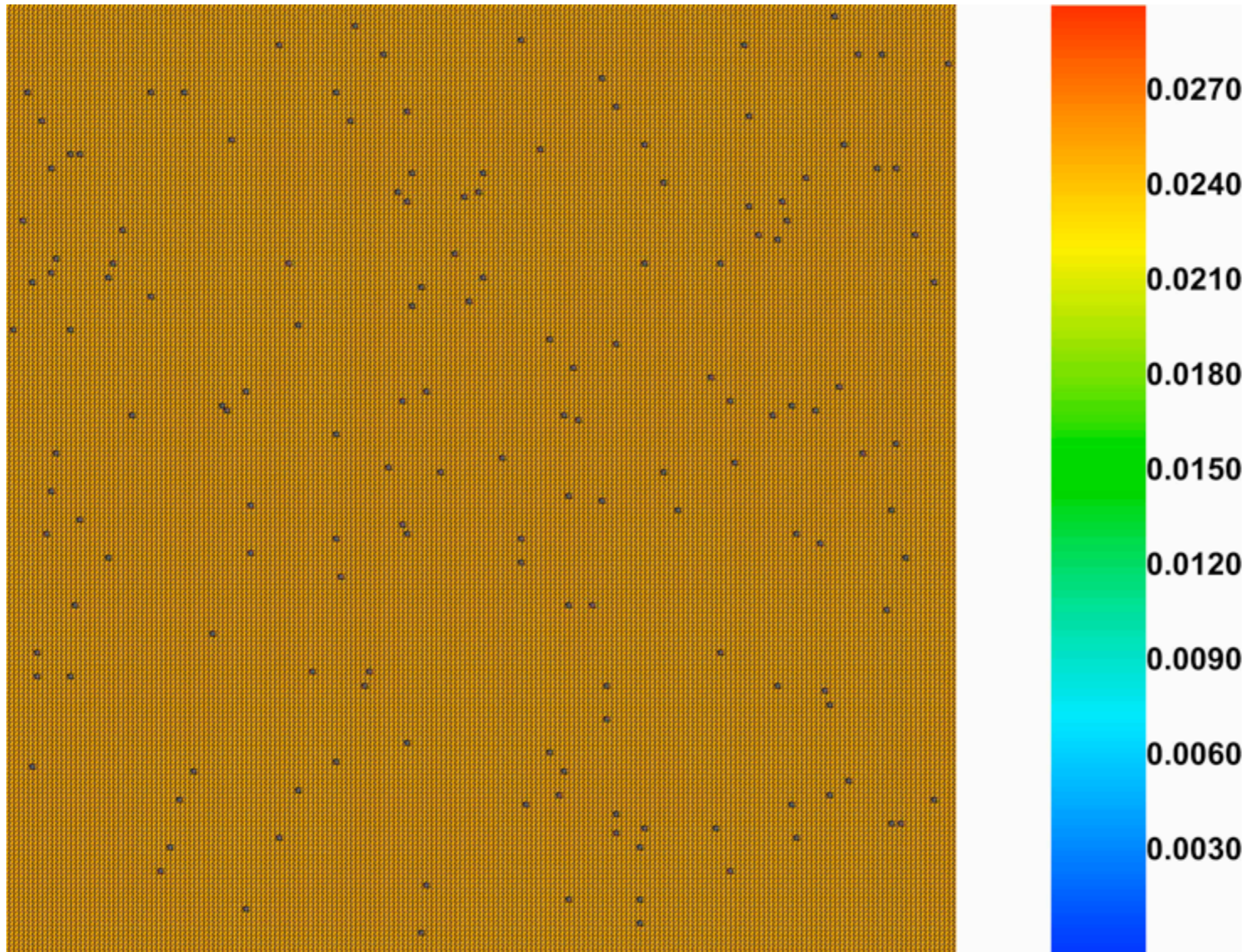
Rens & Merks, Biophys. J. 2017, arXiv:1605.03987

Contractility enhances speed and precision of cell-cell alignment



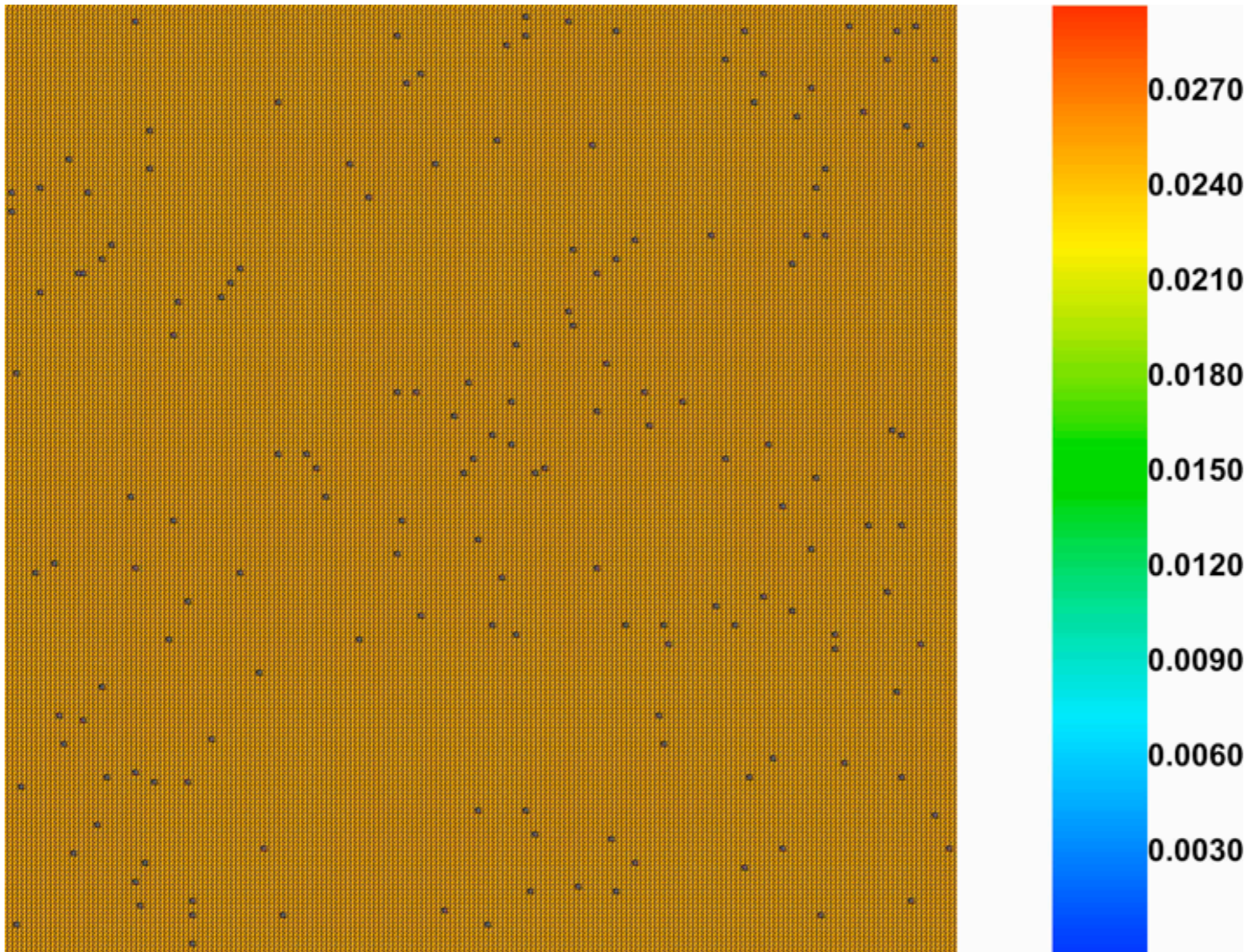
Rens & Merks, Biophys. J. 2017, arXiv:1605.03987

Cells align along stretch orientation... Universiteit Leiden



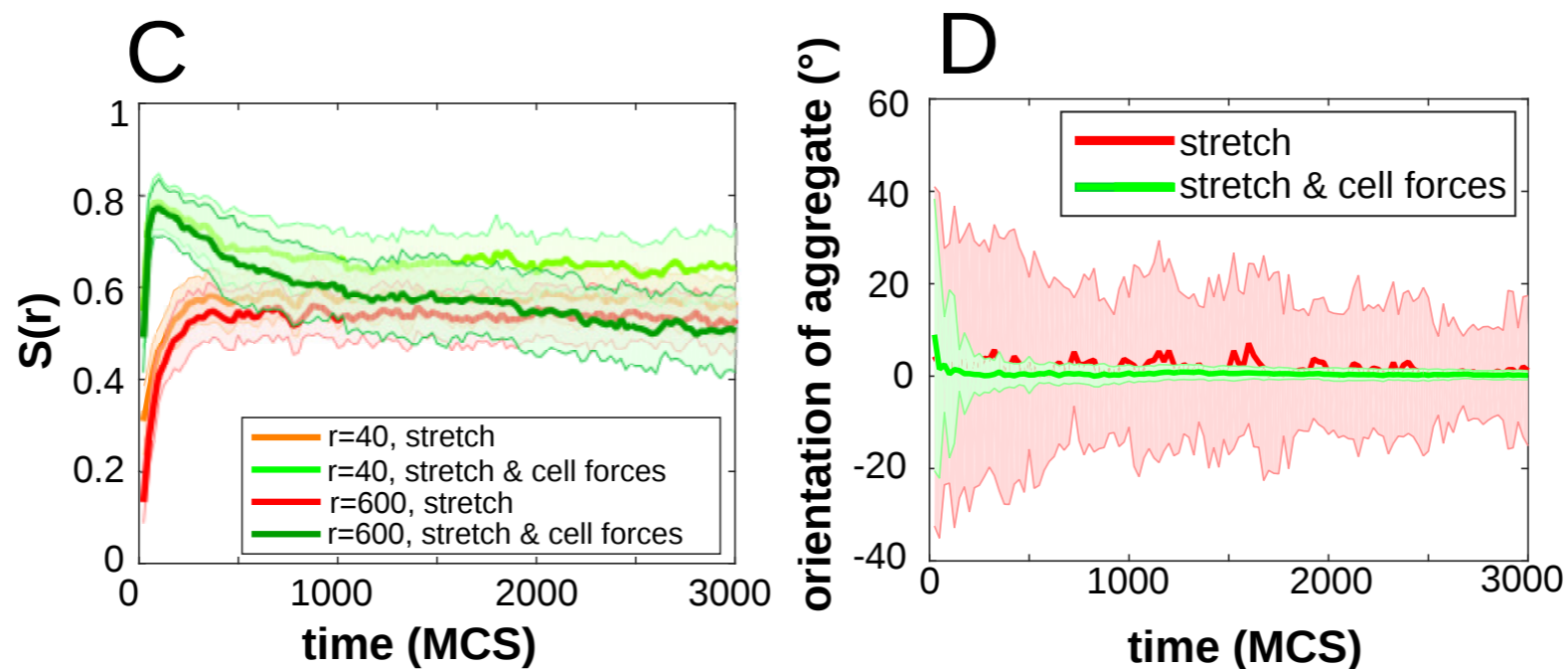
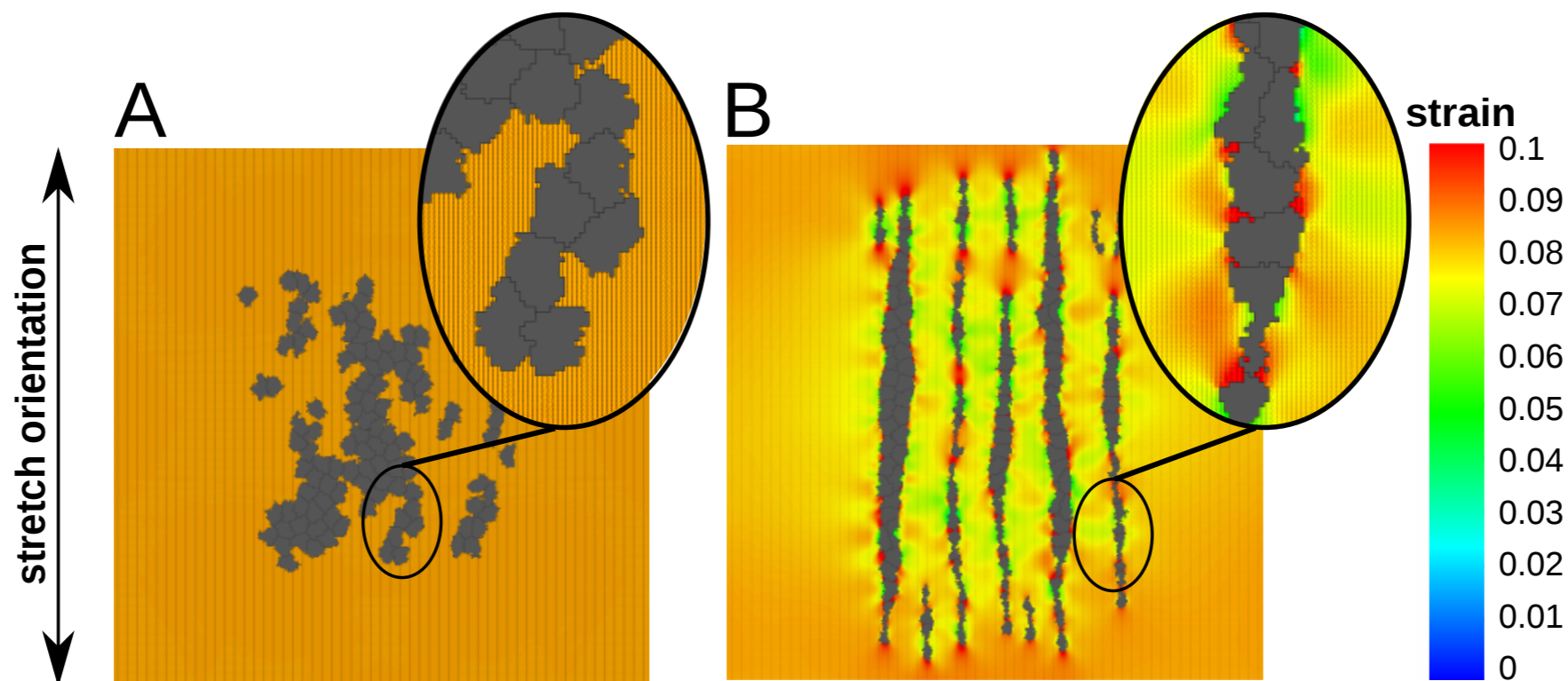
Rens & Merks, Biophys. J. 2017, arXiv:1605.03987

... but only if cells exert forces on matrix



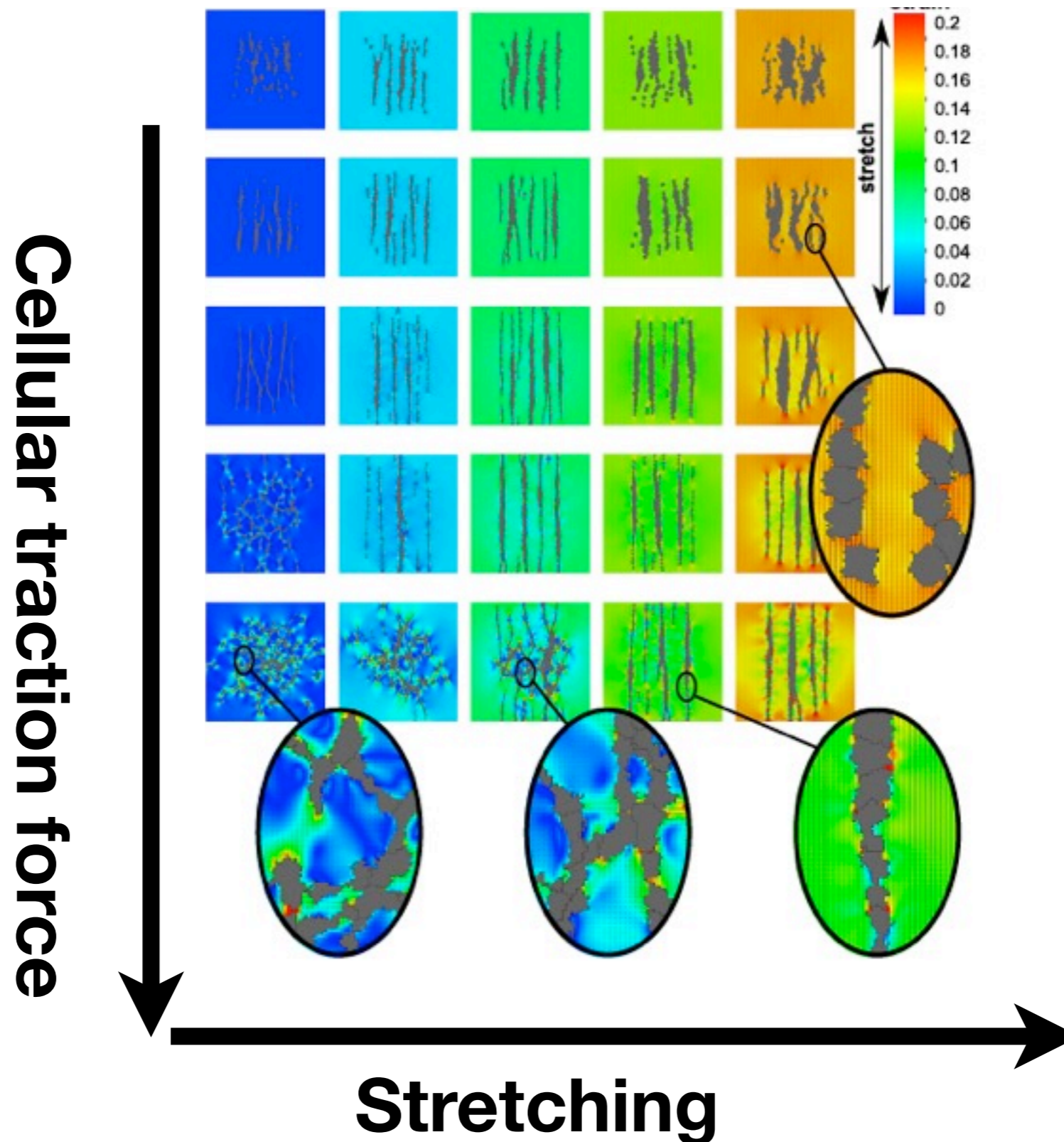
Rens & Merks, Biophys. J. 2017, arXiv:1605.03987

Active cell sensing accelerates response to strain

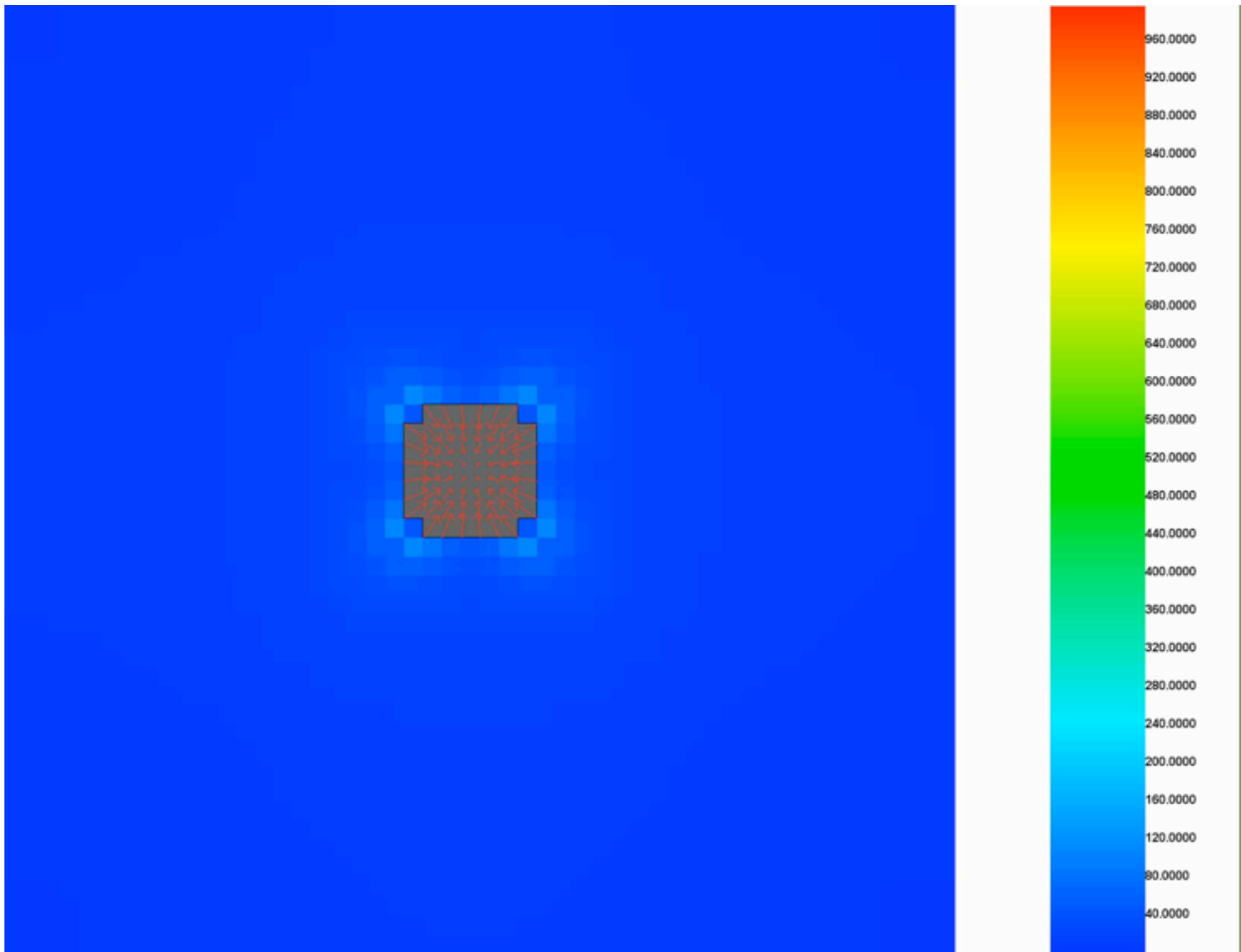


Rens & Merks, Biophys. J. 2017, arXiv:1605.03987

Balance between cell contractility and stretch determines patterning



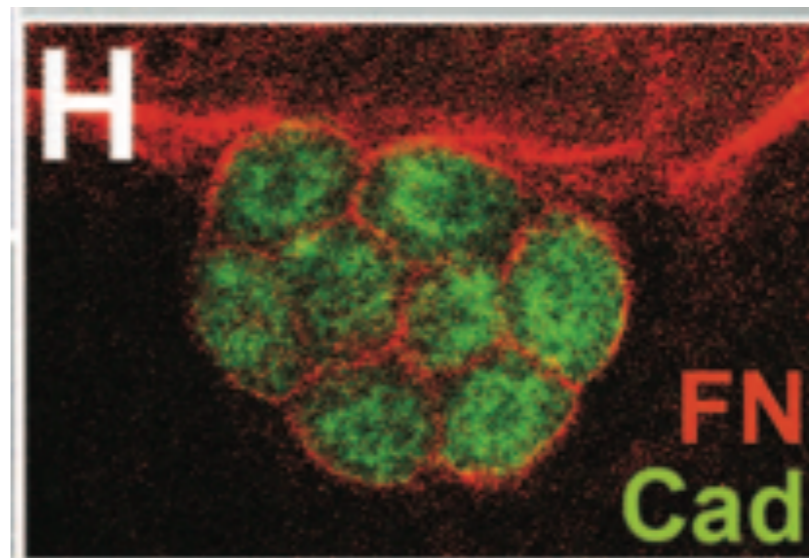
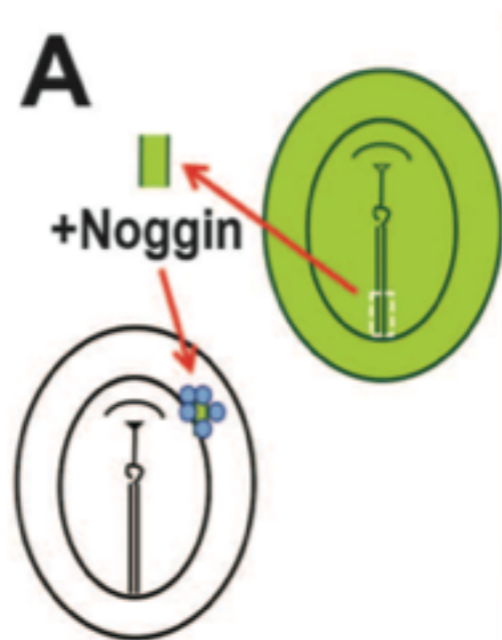
Next step: Focal adhesion kinetics



Lisanne Rens

Mechanical strain during somitogenesis

- “Somites without a clock” (Dias et al. *Science* 2014)
 - Ectopic somites form from dorsalized, ventral primitive streak tissue



- Ectopic somites not polarized, no cyclic gene expression
- **Can extension of body axis put somites in a row?**

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